Associations Between Retinal Pigment Epithelium and Drusen Volume Changes During the Lifecycle of Large Drusenoid Pigment Epithelial Detachments

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PURPOSE. Drusenoid pigment epithelial detachments (PEDs) are a defined path to atrophy in age-related macular degeneration (AMD). We analyzed the relationships between retinal pigment epithelium (RPE) and drusen volume changes during the PED lifecycle, using spectral-domain optical coherence tomography (SD-OCT).

METHODS. Twenty-one cases of drusenoid PED tracked using SD-OCT through periods of growth and collapse were evaluated. Volumetric calculations and piece-wise linear regression analysis were used to determine the breakpoint between growth and collapse. Spectral-domain OCT scans were independently evaluated for the appearance of intraretinal hyperreflective foci, acquired vitelliform lesions (AVLs), and disruptions to the RPEþbasal lamina band. Timing of these events with respect to the breakpoint was statistically evaluated. Morphometric characteristics of drusenoid PEDs were correlated with rate of PED collapse and final visual acuity.

RESULTS. Mean age of subjects was 75.3 years and mean period of follow up was 4.1 years (median 4.5 years; range, 0.6–6.6 years). The lifecycle of drusenoid PEDs was asymmetric, in that the rate of collapse (0.199 mm3/month) is significantly faster (P < 0.001) than the rate of growth (0.022 mm3/month). Appearance of intraretinal hyperreflective foci and AVLs preceded the breakpoint (both P < 0.001). The timing of disruptions to the RPEþbasal lamina band did not differ from the breakpoint (P = 0.510). Maximal height, volume, and diameter of drusenoid PEDs were inversely correlated with final visual acuity (all P < 0.001) and positively correlated with the rate of PED collapse (all P < 0.001).

CONCLUSIONS. Spectral-domain OCT signatures, plausibly attributable to anteriorly migrated RPE and disintegration of the RPE layer, precede or occur simultaneously with changes in volume of drusenoid PED during the lifecycle of this lesion.

Keywords: AMD, drusen, geographic atrophy, pigment epithelial detachment, retinal pigment epithelium

Macular drusen are a defining feature of AMD, a disease that accounts for severe visual loss in a considerable number of people over 65 years of age.1 Drusenoid pigment epithelial detachments (PEDs) are characterized by displacement of the retinal pigment epithelium (RPE) away from Bruch’s membrane (BrM) and are due to enlargement and/or coalescence of soft drusen.2,5 The Age-Related Eye Disease Study defined a PED as a circumscribed, pale or white mound consisting of many large or confluent drusen that was at least 550 μm on the narrowest diameter and appeared elevated on stereoscopic color photographs.3 Roquet et al.,4 also using color photographs, set a size criterion of half-a-disk-diameter. In a recent multimodal imaging review, Mrejen et al.3 defined drusenoid PEDs as structures where the detached RPE has a smooth or undulating contour on spectral-domain optical coherence tomography (SD-OCT) with moderate to high internal reflectivity. Drusenoid PEDs are well-demarcated precursors to atrophy in a subset of AMD patients.5,5 However, the quantitative properties of the PED lifecycle and the cascade of RPE changes that underlie vision loss during this process remain unclear.1,5 Cells of the RPE layer are critically linked to retinal health and disease.5,7 The RPE performs numerous functions essential...
to the photoreceptors and choroid, including phagocytosis of photoreceptor outer segments, absorption of excess light, processing of retinoids for phototransduction, maintenance of the outer blood–retina barrier, and secretion of growth factors, cytokines, and lipoprotein particles.8–11 New information from comprehensive high-resolution histology is prompting a re-examination of RPE fate in the transition to atrophy in AMD.12–15 These recent imaging–histology correlation studies have shown that a range of RPE activities are visible on optimized structural SD-OCT.

A limitation of histologic studies is that they provide structural information from only a single time-point in the disease course, yet such studies can facilitate use of SD-OCT information from clinic populations to study RPE changes as a function of time. The current report quantifies the timeline of drusenoid PED growth and collapse using high-resolution SD-OCT imaging. Retinal pigment epithelial changes that are associated with volumetric alterations during the PED lifecycle are also defined. Finally, this report identifies morphometric properties of drusenoid PEDs that are significant prognosticators of poor final visual acuity.

METHODS

This study followed the Tenets of the Declaration of Helsinki and was approved by the Western institutional review board.

Patient Inclusion Criteria

Large drusenoid PEDs that were judged to be morphometrically similar to a forthcoming imaging–histology correlation of a donor eye (maximum diameter of base, 1916 μm, and maximum height of dome, 432 μm) were included in this study (illustrative case shown in Fig. 1). Another reason for evaluating large drusenoid PEDs in this study is because they are straightforward to track on OCT and demonstrate highly reproducible volumetric measurements (as described below). Only cases where SD-OCT imaging data was available from the baseline visit (first clinic visit) to the time of PED collapse were included for quantitative analysis in this study. Cases were selected in a retrospective manner from a consecutive series of PEDs seen between June 2015 and January 2016 by two retina specialists (LAY and KBF) at the Vitreous Retina Macula Consultants of New York. In those patients with bilateral drusenoid PEDs, a single eye was selected at random for inclusion into this study. Exclusion criteria included: (1) choroidal neovascularization confirmed with SD-OCT and dye angiography (fluorescein or indocyanine green), (2) retinal artery/vein occlusion, diabetic retinopathy, previous vitreoretinal surgery, and pathologic myopia, (3) geographic atrophy at baseline visit, (4) prior ocular therapies such as laser photocoagulation or intravitreal therapy, (5) PEDs due to other causes as central serous chorioretinopathy and polypoidal choroidal vasculopathy, (6) inability to visualize BrM on SD-OCT, and (7) fewer than six OCT volume scans performed between baseline visit to the time of PED collapse.

Imaging

Color photography, fluorescein angiography (FA), and indocyanine green angiography (ICGA) images were acquired using the Topcon TRC-50XF fundus camera (Topcon Medical Systems, Paramus, NJ, USA). Fundus autofluorescence (FAF) images were obtained using the Spectralis HRA + OCT (Heidelberg Engineering, Heidelberg, Germany) or the Topcon TRC-50XF fundus camera (Topcon Medical Systems), respectively. All eyes underwent SD-OCT imaging using the Spectralis OCT (Heidelberg Engineering) at each visit. Eye tracking and image registration functions were enabled for all image acquisitions. The SD-OCT scan protocol for the cohort is summarized in Table 1.

SD-OCT Definitions for Outer Retinal Bands and Drusenoid Pigment Epithelial Detachment

In a healthy eye the fourth outer retinal hyperreflective band includes the RPE and BrM.16,17 In a PED, by definition the RPE is separated from BrM and therefore it is incorrect to call the hyperreflective band that anteriorly delimits a PED the RPE-BrM. When the RPE is separated from BrM in the course of histologic preparation, either a basal lamina of 0.15-μm thickness or basal laminar deposit (BLamD) of variable thickness adheres to the cells and not to BrM, suggesting a natural cleavage plane between the RPE basal lamina and the inner collagenous layer of BrM.18 In this study, we use the designation RPE-basal laminar (BL) to indicate the fourth outer retinal hyperreflective band in drusenoid PED.

Quantitative Volumetric Analysis of Drusenoid Pigment Epithelial Detachments

The PED volume at each visit was determined by applying the Cavalieri principle of stereological analysis to SD-OCT volumetric data (Fig. 2).19 All measurements were attained using a 1:1 micron scaled image on SD-OCT software. Pigment epithelial detachment volume was determined using the following steps: (1) the area between the outer RPE+BL boundary and inner boundary of BrM for each slice in the PED volume scan was calculated using manual planimetry, (2) the volume between two adjacent OCT slices (herein called a segment) was then determined using this equation:

$$d \left( \frac{A_x + A_{x+1}}{2} \right)$$

where: d = distance between adjacent slices in μm, A = Area between the RPE+BL and BrM in μm², and x = OCT slice number.

(3) Total PED volume was calculated by summing the volumes of individual segments. The number of segments in the OCT volume was (n – 1), where n = total number of slices that spanned the PED. Plots of PED volume versus time were then generated (Fig. 3). For eight eyes, volumetric measurements were repeated three times, on different days, and the...
coefficient of variation was determined to measure the reproducibility of manual tracings. For these eyes, the mean of the three measurements was used for plots of PED volume versus time.

The following measurements were also attained at the visit where the volume of the PED was calculated to be greatest (Fig. 2):

1. Maximal diameter: defined as the greatest PED base diameter using BrM as the reference plane.
2. Maximal height: defined as the greatest distance between BrM and the outer boundary of the RPE+BL.
3. Maximal angle between RPE+BL and BrM: defined as the greatest angle within the entire volume scan, on both nasal and temporal aspects.

Pigment epithelial detachment area, height, and diameter measurements were determined using the contained software (Spectralis Viewing Module 4.0.0.0; Heidelberg Engineering), as was baseline central subfoveal choroidal thickness. Angle measurements were determined using Image J (version 1.43; http://imagej.nih.gov/ij/; provided in the public domain by the National Institutes of Health, Bethesda, MD, USA) following exportation of tif files.

Qualitative Evaluation of Pigment Epithelial Detachment and Outer Retinal Changes

Spectral-domain OCT volumetric scans were analyzed in a masked fashion for the following RPE structural changes:

1. Intraretinal hyperreflective foci: our recent histology-OCT correlation study demonstrated that intraretinal hyperreflective foci overlying drusenoid PEDs correspond to RPE cells, either singly or in groups. The time point at which intraretinal hyperreflective foci on SD-OCT were first observed was recorded.
2. Disruptions to the RPE+BL band with associated increased light transmission (hypertransmission) through the PED into the choroid. Hypertransmission below the RPE is considered a reliable indicator of absence of the RPE layer, in geographic atrophy. The time point at which disruptions to the RPE+BL band first occurred and their location in the PED dome (apex or...
A single SD-OCT B-scan (A) is used to illustrate the measurements that were acquired from the PED, which included maximum height and diameter. The angle of intersection between the RPE+BL band and BrM in the temporal (α) and nasal (β) margins of the PED was also determined. Manual tracing of Bruch’s membrane and the outer margins of the RPE+BL band was used to calculate the cross-sectional area (blue shade) of the drusenoid PED. Area measurements from each slice in the OCT volume scan were then used to perform volumetric calculations (B). Three contiguous OCT slices illustrate the application of the Cavalieri principle in stereology for calculating volume. The volume of a segment, defined as the volume between two contiguous OCT slices, was determined by averaging the area of the drusenoid PED in the two slices and multiplying it by the distance (d) separating the slices. Summation of volumetric measurements from each segment allowed calculation of the entire volume of the PED.
compared using a 2-sample t-test. T-tests were also used to compare the time of first occurrence of intraretinal hyperreflective foci, the time of first occurrence of RPE + BL band disruption, the time of first occurrence of AVLs, and the time of slope changes on the PED volume curve. Pearson’s or Spearman’s correlation was used to evaluate relationships between PED measurements including maximal height, maximal diameter, maximal volume, temporal angle, nasal angle, baseline visual acuity, and the rate of drusen volume decrease and logMAR final visual acuity. A P value less than 0.050 was considered significant. Quantitative results are expressed as mean ± SEM.

RESULTS

General

A total of 216 eyes from 121 patients were reviewed of which 21 eyes from 21 patients met the inclusion criteria for this study. All eyes were graded as intermediate AMD at baseline visit. An illustrative case of a study eye is presented in Figure 1. On ophthalmoscopy, drusenoid PEDs appeared as yellow lesions with smooth or scalloped borders. Areas of RPE hyperpigmentation were seen on the PED surfaces, and these regions typically appeared hyperautofluorescent on FAF imaging. All patients were imaged with FA, and 10 patients were also imaged with ICGA. Fluorescein angiography demonstrated hyperfluorescence at the site of the PED that was more intense in the late stages of the angiogram, but did not extend beyond its margins. Areas of RPE hyperpigmentation appeared hypofluorescent on FA. On ICGA there was obscuration of the choroidal vessels at the site of the PED and hypofluorescence at sites of PED hyperpigmentation.

Mean age of subjects was 75.3 ± 8.7 years (range, 56–91 years). The cohort comprised of 16 females, 5 males, 9 right eyes, and 12 left eyes. Mean time-period between the first OCT scan and last OCT scan was 4.1 ± 1.7 years (median 4.5 years, range, 0.6–6.6 years). Mean best-corrected visual acuity at baseline visit was 0.22 logMAR (Snellen equivalent 20/33; range, 0.1–0.5 logMAR). Best-corrected visual acuity at final visit was higher on the logMAR scale, 0.8 logMAR (Snellen equivalent 20/127; range, 0–2.3 logMAR; P < 0.001). Mean subfoveal choroidal thickness was 216.3 ± 87.9 µm (Table 2).

Drusenoid PED Volume Changes Over Time

An illustrative plot of PED volume against time is provided in Figure 3. Mean coefficient of variation for repeated PED volume measurements from eight eyes was 0.14 ± 0.04. All PEDs demonstrated dynamic changes characterized by a phase where PED volume gradually increased followed by a period of rapid reduction in volume to a floor value of zero or almost zero (Fig. 3A). At the final visit, the PED volume was measured as 0 mm³ in 16 of 21 eyes (76%) and less than 10% of maximal PED volume in the remaining five eyes. Piece-wise linear regression analysis identified a breakpoint where the slope changed significantly between periods of growth and collapse in all cases (Fig. 3A). The absolute value of the slope during PED collapse (0.199 ± 0.176 mm³/month) was significantly greater than the absolute value of the slope during PED growth.

### Table 2. Summary of Morphometric Dimensions of Drusenoid Pigment Epithelial Detachment and Subfoveal Choroidal Thickness

<table>
<thead>
<tr>
<th>Measured Parameters</th>
<th>Mean ± SEM</th>
<th>Median</th>
<th>Minimum</th>
<th>Maximum</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maximal diameter, µm</td>
<td>2426.5 ± 1096.2</td>
<td>2525</td>
<td>904</td>
<td>4691</td>
</tr>
<tr>
<td>Maximal height, µm</td>
<td>352.9 ± 156.3</td>
<td>338</td>
<td>164</td>
<td>659</td>
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<tr>
<td>Maximal volume, mm³</td>
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<td>0.8</td>
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<td>6.8</td>
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<tr>
<td>Nasal angle, °</td>
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<td>37.9</td>
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<td>71</td>
</tr>
<tr>
<td>Temporal Angle, °</td>
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<td>43.3</td>
<td>23.4</td>
<td>65.2</td>
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<tr>
<td>Subfoveal choroidal thickness, µm</td>
<td>212.7 ± 88.7</td>
<td>219</td>
<td>52</td>
<td>376</td>
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</table>
(0.022 ± 0.028 mm³/month; \( P < 0.001 \); Fig. 3B). The mean period of time from baseline visit to the breakpoint of the PED drusen lifecycle was 23.5 ± 19.9 months (range, 3–61 months). The mean period of time from breakpoint to complete collapse of the PED was 10.4 ± 4.2 months (range, 5–18 months). Visual acuity between baseline and the visit of maximal PED volume was not significantly different (\( P = 0.105 \)). Visual acuity at the final visit was significantly worse than baseline visit and the visit where PED volume was maximal (both \( P \leq 0.021 \)).

**RPE Changes During the Lifecycle of Drusenoid PEDs**

The relationships between changes to the RPE-BL band, as seen on SD-OCT, and volume changes during the drusenoid PED lifecycle are illustrated in Figure 4. Intraretinal hyperreflective foci were identified in all eyes (mean time of 4.1 ± 15.4 months from baseline visit). AVLs were seen in 66.7% of eyes (3.8 ± 13.4 months from baseline visit) and disruption of the RPE+BL band was seen in 81.0% of eyes (230.9 ± 106.3 months from baseline visit). The time point at which intraretinal hyperreflective foci were first observed did not differ from the time point when AVLs were first seen (\( P = 0.950 \)). The time points at which intraretinal hyperreflective foci and AVLs were first observed was significantly earlier than the time point where disruption of the RPE+BL was first observed (\( P = 0.004 \) and \( P = 0.005 \), respectively).

Intraretinal hyperreflective foci were seen on average 19.4 ± 17.0 months before the breakpoint of the drusenoid PED lifecycle (\( P = 0.001 \)). Acquired vitelliform lesions occurred on average 15.6 ± 15.9 months before the breakpoint of the drusenoid PED lifecycle (\( P = 0.002 \)). Disruptions in the RPE+BL band were first observed a mean period of 3.7 ± 10.4 months before the breakpoint of the drusenoid PED lifecycle but this difference was not significant (\( P = 0.510 \)).

Of the eyes with disruptions to the RPE+BL band, 76.5% of cases demonstrated disruptions at the apex of the PED only, while 23.5% demonstrated disruptions to the apex and base of the PED (Fig. 5). The mean length of RPE+BL disruptions was 230.9 ± 106.3 μm (range, 94–434 μm). There was no evidence of lateral or anterior retraction of the RPE adjacent to the area of RPE+BL defect. Disruptions were associated with hyper-transmission below the RPE (Fig. 5).

**Correlations Between PED Morphometry, Final Visual Acuity, and Rate of Collapse**

Maximal PED morphometric measurements for each case are summarized in Table 2. Shapiro-Wilk normality testing demonstrated that data for maximal PED height and volume were not normally distributed (both \( P \leq 0.015 \)) while maximal PED diameter was normally distributed (\( P = 0.076 \)). Maximal PED diameter (\( P < 0.001 \); \( r = -0.767 \)), maximal PED height (\( P < 0.001 \); \( r = -0.724 \)), and maximal PED volume (\( P < 0.001 \); \( r = -0.866 \)) were positively correlated with the rate of drusenoid PED collapse (Fig. 6). Baseline visual acuity (\( P = 0.356 \)), nasal angle of PED (\( P = 0.641 \); \( r = -0.111 \)), and temporal angle of PED (\( P = 0.445 \); \( r = -0.182 \)) were not correlated with the rate of drusenoid PED collapse.

Maximal PED diameter (\( P < 0.001 \); \( r = 0.685 \)), maximal PED height (\( P < 0.001 \); \( r = 0.601 \)), and maximal PED volume (\( P < 0.001 \); \( r = 0.725 \)) were inversely correlated with final visual acuity (Fig. 7). Baseline visual acuity (\( P = 0.851 \)), nasal angle of PED (\( P = 0.268 \)), and temporal angle of PED (\( P = 0.294 \)) were not correlated with final visual acuity (Fig. 7).
DISCUSSION

Drusenoid PEDs are a recognized phenotypic manifestation of nonneovascular AMD that are associated with increased risk of progression to geographic atrophy.\(^4,5\) Understanding the temporal properties of the PED lifecycle is therefore important as it may identify biomarkers that can be used as clinical trial endpoints to guide treatment and prevent irreversible visual loss. This study quantified the lifecycle of large drusenoid PEDs based on longitudinal follow-up with structural SD-OCT. Piecewise linear regression analysis revealed that the growth and collapse phases of the drusenoid PED lifecycle are asymmetrical; the rate of drusenoid PED collapse (mean 0.2 mm\(^3\)/month) was significantly faster than the rate of PED growth (mean 0.02 mm\(^3\)/month). Important determinants of the rate of drusenoid PED collapse included maximal PED volume, height, and diameter. Larger drusenoid PEDs demonstrated a faster rate of collapse. Recently, Schlanitz and colleagues\(^{25}\) evaluated drusen volume changes using SD-OCT and polarization-sensitive OCT in eyes with drusen of AREDS categories 2 and 3. These authors showed that the drusen lifecycle was also comprised of periods of growth and regression. The relationships between RPE and drusen volume changes were not explored in that study, however. Collectively, the findings of their study and this report demonstrate a degree of overlap in the natural history of drusen and drusenoid PEDs.

**Figure 5.** Spectrum of RPE-BL disruptions during the drusenoid PED lifecycle. Spectral-domain OCT images from different patients (A-E) demonstrate areas of RPE-BL disruption (red arrows) with hypertransmission to the choroid. Patients A, B, C and E demonstrate disruptions at the apex of the PED while Patient D demonstrates a disruption at the base of the PED.
Structural and functional changes to RPE cells are inherently linked to the pathophysiological cascades underlying atrophy and photoreceptor loss due to AMD. Intraretinal hyperreflective foci are frequently seen on SD-OCT in eyes with AMD, yet there has been some controversy regarding their cellular origin. In 2006, Peirroni et al. proposed that hyperreflective foci overlying drusenoid PEDs signified RPE migration into the retina. Framme and colleagues proposed that these features in AMD eyes reflect leucocytes or migrating RPE cells. High-resolution studies of eyes with AMD recently completed by Curcio and colleagues, including two cases with large subfoveal serous and hemorrhagic PED, established a histologic basis for RPE morphology visible in SD-OCT. These histopathology-OCT correlations demonstrated that hyperreflective intraretinal foci represent fully pigmented and nucleated RPE cells apparently migrating in an anterior direction, and focal thickening of the RPEþBL band resembling AVLs represent subretinal, extracellular plaques of numerous RPE granules, and some outer segment debris.

The present study delineated the time course of RPE changes during the drusenoid PED lifecycle using these previously determined OCT-histologic correlations. All eyes in our cohort demonstrated one or more RPE structural alterations prior to PED collapse including appearance of intraretinal hyperreflective foci, formation of AVLs, and/or disruption of the RPEþBL band. Retinal pigment epithelial changes that were identified in this study may represent distinct manifestations of different RPE degenerative pathways (as hypothesized) or different manifestations of a time-dependent continuum. Mrejen et al. proposed that RPE atrophy in drusenoid PEDs was due to prolonged displacement from the choriocapillaris, its major source of nutrition. This may be one reason why disruptions to the RPEþBL band were frequently seen at the apex of the PED, the point of greatest separation from the choriocapillaris. It is important to distinguish disruptions to the RPEþBL band in drusenoid PEDs from RPE rips, which are a recognized complication of neovascular AMD. Retinal pigment epithelial tears in neovascular AMD are due to abnormal tractional forces acting on the PED and typically occur at the margin, which is the point of greatest mechanical stress in a dome-shaped structure. Unlike RPE tears due to neovascular AMD, ripping or retraction of the RPE was not seen at sites of RPEþBL disruption in this study. Our findings therefore suggest that RPE disruptions in drusenoid PEDs are principally due to activities intrinsic to RPE cells such as necroptosis, apoptosis, and migration, in response to environmental signals, rather than mechanical forces. Ouyang and colleagues performed longitudinal evaluations of drusenoid lesions and demonstrated that intraretinal hyperreflective lesions increased the risk of progression to atrophy. The findings of our study support this conclusion. However, we emphasize that anterior migration of RPE cells was not predictive of poor visual acuity in all eyes as some of the cases in our cohort retained a vision of 20/20.

**Figure 6.** Predictors of final visual acuity (VA) following drusenoid PED collapse. Scatter plots demonstrate that PED diameter (A), PED height (B), and PED volume (C) are significantly correlated with final visual acuity (all P < 0.001). Baseline VA (D), nasal angle of PED (E), and temporal angle of PED (F) were not correlated to final VA. The best-fit trend line and R² are presented on each plot.
following collapse of the PED. Pathogenic mechanisms beyond anterior migration of RPE cells are therefore contributory to the development of geographic atrophy in AMD.

As RPE-derived molecules and multimolecular complexes are major druse components, it is plausible that dysfunction or loss of RPE cells is associated with a decline in drusen volume. Multiple lines of evidence suggest that soft drusen and basal linear deposit form because large cholesterol-rich lipoproteins containing apolipoproteins B and E are secreted from the basal surface of the RPE and accumulate throughout adulthood on the inner collagenous layer of aging BrM. In our study, the breakpoint in the drusenoid PED life cycle coincided with the time where RPE disruptions were observed. Thus, the presence of drusen may be a positive sign that RPE is healthy enough to continue secreting material and thus maintain the lesions. Sarks et al. observed that RPE attenuation, as seen on FA and histology, was associated with the early stages of drusen regression. However, their report did not delineate temporal relationships between the RPE and drusenoid PED volume changes. Microglia and activated Müller cells have all been identified in lesion debris of atrophic AMD eyes and in eyes treated by nanosecond laser for drusen abatement. These cells may also share responsibility for clearing. Future research is needed to determine if PED collapse and loss of drusenoid material is due to reduced secretion by RPE, active removal of drusen constituents by scavenging cells, or both.

The inclusion criteria for this study ensured that we evaluated a population of PEDs rather than just large drusen. A considerable advantage with evaluating large drusenoid PEDs is fewer errors associated with image segmentation, particularly with respect to defining the boundary between the RPE and drusen. Previous studies have shown that the boundaries of the RPE layer can be obscured on OCT due to noise and poor image quality, thus resulting in inaccuracies in drusen volume calculations. The coefficient of variation for repeat measurements in this study was low (0.14 ± 0.04), suggesting that the methodology for volumetric analysis was reproducible. The inclusion criteria used in this study ensured that we evaluated a population of PEDs rather than just large drusen. A considerable advantage with evaluating large drusenoid PEDs is fewer errors associated with image segmentation, particularly with respect to defining the boundary between the RPE and drusen.
study, however, limits the degree to which these findings can be extrapolated to other pathways leading to atrophy, such as soft drusen in the absence of PED. Other limitations of this study include its limited sample size and the nonstandardized OCT imaging protocol. Nevertheless, our study represents the first attempt to link cellular-level changes in the RPE layer to the dynamism of drusenoid PED.

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References


