Association of Glaucoma-Susceptible Genes to Regional Circumpapillary Retinal Nerve Fiber Layer Thickness and Visual Field Defects

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See the appendix for the members of the Nagahama Study Group.

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PURPOSE. To examine the associations of the earlier reported glaucoma-related genes to the regional circumpapillary retinal nerve fiber layer thicknesses (cpRNFLTs) and corresponding visual field defects.

METHODS. We studied 756 patients with primary open-angle glaucoma (POAG) and 3094 normal controls. Each participant was genotyped for nine single nucleotide polymorphisms (SNPs) of four glaucoma-susceptible genes: the CDKN2B, TMCO1, CAV1/CAV2, and SIX1/SIX6 genes. For the SNPs that were significantly associated with the POAG case-control analyses, the associations of SNP genotypes with the cpRNFLTs of 12 sectors were also analyzed, and then finer assessments were performed using 768 points of the cpRNFLT and corresponding visual field defect sensitivities using case-only subjects.

RESULTS. We confirmed that there was a significant association of the CDKN2B gene to POAG. For the suggested region-specific associations of these genes with the 12-sector cpRNFLT, a 768-point cpRNFLT examination showed that rs4977756 near CDKN2B had significant signal peaks in the temporal region at 330° to 360° and 0° to 30° (maximum $P = 2.92, 0.05$). These region-specific signals were validated by the corresponding visual field defect patterns of the paracentral/lower hemifield ($P < 0.05$).

CONCLUSIONS. Genetic association analyses using the cpRNFLT with 768 points suggest that the CDKN2B gene was associated with paracentral/lower hemifield scotomas. Our regional association analyses on cpRNFLT allow detailed characterization of glaucoma-related genes and should be a new target for genomic studies for glaucoma endophenotypes.

Keywords: glaucoma, retinal nerve fiber layer thickness, CDKN2B, visual field defect, genetics

Glaucoma

Glaucoma is a complex vision-threatening disorder that is caused by a combination of genetic and environmental factors. It is one of the highest causes of acquired blindness worldwide. Several genome-wide association studies (GWAS) have identified glaucoma susceptibility genes including CDKN2B (AS1), CAV1/CAV2, TMCO1, and SIX1/SIX6. Some of these genes have been confirmed to be associated with glaucoma or related endophenotypes, such as the vertical cup-to-disc ratio (VCDR) and intraocular pressure (IOP).

Glaucoma is characterized by glaucomatous optic neuropathy (GON) with corresponding visual field defects (VFDs) that result from the death of retinal ganglion cells (RGCs) and their axons. Because the VFDs are the most important parameter for GON diagnosis, the relationship between the glaucoma susceptibility genes and the areas of the VFDs has been extensively studied. However, visual field tests depend on subjective responses of the glaucoma suspects, and the commonly used standard automated perimetry (SAP) 30-2 program examines less than one-sixth of the whole visual field. Thus, another parameter for GON evaluations is needed to determine the relationship between the glaucoma susceptibility genes and the VFDs.

Recent advances in optical coherence tomography (OCT) have allowed clinicians and researchers to make quantitative evaluations of the circumpapillary retinal nerve fiber layer (cpRNFL) thicknesses. Because the degeneration of the RGCs and their axons usually precedes the depression of visual function, assessments of the cpRNFLT (cpRNFL) should be a better way to detect the early changes of GON with a high degree of reproducibility.

Earlier genetic studies evaluated the cpRNFLT by dividing it into four to six sectors, and polymorphisms in the SIX1/SIX6 gene were found to be significantly associated with cpRNFL thinning in the superior and inferior regions but not in the nasal and temporal regions. The question then arises whether analysis of the cpRNFL with finer sectors might provide more detailed information on the regional changes of the GON.
Thus, the purpose of this study was to determine whether the glaucoma susceptibility genes identified in earlier studies were significantly associated with primary open-angle glaucoma (POAG) in Japanese patients. In addition, we examined whether these glaucoma susceptibility genes were significantly associated with alterations of the 12 sectors of the cpRNFLT along the OCT circle scans. The region-specific associations of the genes to GON were further examined at 768 points cpRNFLT and confirmed by the corresponding VFDs.

Methods

Ethics Statement

All procedures used in this cross-sectional observational study adhered to the tenets of the Declaration of Helsinki. The Institutional Review Board and Ethics Committee of Kyoto University Graduate School and the Faculty of Medicine Ethics Committee, the Ad Hoc Review Board of the Nagahama Cohort Project, and the Nagahama Municipal Review Board of Personal Information Protection approved the protocols of this study. All patients were fully informed on the purpose of and procedures to be used in this study, and a written informed consent was obtained.

Study Design

First, we performed a case-control replication analysis on the nine single nucleotide polymorphisms (SNPs) of four glaucoma susceptibility genes, namely, CDKN2B(AS1), TMCO1, CAV1/CAV2, and SIX1/SIX6, in POAG patients and control subjects. Second, we determined whether there were significant associations between the SNP genotypes and any of the 12 sectors of the cpRNFLT along the OCT circle scan. The region-specific associations of the genes to GON were further examined at 768 points cpRNFLT and confirmed by the corresponding VFDs. The associations with the corresponding VFDs were also analyzed to validate these associations.

Subjects

All case subjects were Japanese who were diagnosed with POAG in the Department of Ophthalmology and Visual Sciences of the Kyoto University Hospital. All patients who were examined between January 2008 and October 2014 and consented to have their genomic DNA analyzed were included. The subjects had a complete ophthalmic examination including an objective determination of the refractive error (spherical equivalent), keratometry, IOP measurements with a Goldmann applanation tonometer, slit-lamp biomicroscopy, gonioscopy, indirect ophthalmoscopy, axial length measurements by partial coherence interferometry (IOL Master; Carl Zeiss Meditec, Inc., Dublin, CA, USA), SAP (Humphrey Visual Field Analyzer [HFA]; Carl Zeiss Meditec, Inc.), and cpRNFLT examinations with the Spectralis OCT (Heidelberg Engineering GmbH, Heidelberg, Germany). The exclusion criteria were prior intraocular surgery except for cataract or glaucoma surgery, high myopia with an axial length ≥26 mm, and presence of other ocular diseases that can influence the visual fields in the studied eye. In the end, 756 patients with POAG were studied.

We used the data of 3094 healthy Japanese volunteers who were enrolled in the Nagahama Prospective Genome Cohort for the Comprehensive Human Bioscience dataset (The Nagahama Study) for the control values. This community-based prospective multi-omics cohort study has been described in detail elsewhere.22-23 Genome-wide SNP information was available for all subjects. The control subjects also had objectively determined ophthalmic data: the refractive errors, keratometry, fundus photographs, and axial length measurements.24 We included all participants with axial length <26 mm because the visual field information was not available for them.

Diagnosis of Primary Open-Angle Glaucoma

POAG was diagnosed by the presence of pathognomonic VFDs that are consistent with GON. GON was diagnosed by the glaucomatous appearance of the optic nerve head, for example, diffuse or localized rim thinning. A glaucomatous VFD was defined as one having the following SAP results: (1) outside the normal limits on glaucoma hemifield tests; (2) three abnormal points with P < 5% probability of being normal; one
with \( P < 1\% \) by pattern deviation; or (3) pattern standard deviation of \( 5\% \) if the visual field was otherwise normal. An open angle was defined as one not having peripheral anterior synechia or appositional closure of the angle as determined by gonioscopy or anterior segment OCT. The eyes with secondarily elevated IOPs, such as pseudoexfoliation syndrome or uveitis, were excluded.

**Circumpapillary Retinal Nerve Fiber Layer Thickness**

The Spectralis OCT was used to obtain circular B-scans of 12° diameter centered on the optic disc, that is, a circumpapillary scan, and each B-scan was obtained by averaging 16 images to reduce speckle noise. The cpRNFL thickness was measured as the distance between the inner border of the internal limiting membrane and the outer border of the RNFL semiautomatically with the built-in software.

The RNFL thickness values at 768 points along the 360° OCT circle scan were obtained by the RNFL Export software (Heidelberg Engineering Gmbh). All of the acquired images of Spectralis OCT acquisition module version 5 or later (from September 2009) were included. We excluded eyes with extensive peripapillary atrophy (larger than 3.46 mm in diameter or affecting the cpRNFL scans), RNFL schisis, Spectralis quality index < 5 dB, or other segmentation errors of the RNFL in the OCT images. The left-eye data were converted into a right-eye format. We divided the 768-point cpRNFLT values into 12 sectors for the association analyses between the glaucoma-susceptible genes and regional cpRNFLT (Fig. 1).

**Visual Field Sensitivity**

Reliable visual field tests were those with fixation loss of 20% or less, false-positive rates of 15% or less, and false-negative rates of 15% or less in the 24-2/30-2/10-2 SAP program. For perimetric analyses, all acquired data of the patients were analyzed, but eyes with other ocular diseases or neurologic diseases that can influence visual fields were excluded. We analyzed the visual field sensitivities of paracentral 10° or 24° and the upper and lower hemifields using HFA 10-2 or HFA 24-2/30-2 Swedish Interactive Thresholding Algorithm (SITA) standard. The total deviation values on a decibel scale were converted to 1/Lambert (1/L) scale at single test positions with the following formula:

\[
\text{dB} = 10 \times \log_{10}(1/L)
\]

Then, the average of 1/L scales at 68 or 52 test points in the HFA 10-2 or 24-2/30-2 programs was analyzed, respectively. Those of the 34 or 26 test points in each hemifield were averaged to evaluate the upper and lower hemifields.

**SNP Selection and Genotyping**

Previously, nine SNPs for \( CDKN2B(AS1) \), three SNPs for \( CAV1/CAV2 \), two SNPs for \( TMCO1 \), and one SNP for \( SIX1/SIX6 \) had genome-wide significant associations with POAG in both Asians and Caucasians. We calculated the linkage disequilibrium (LD) between each associated SNP with the HapMap dataset of JPT/CHB using the SNAP software (http://www.broadinstitute.org/mpg/snap/ldsearch.php; in the public domain), and selected representative SNPs from the LD blocks with \( R^2 > 0.8 \). Minor allele frequencies (MAF) and reported odds ratio (OR) were also considered to exclude SNPs with insufficient statistical power (<0.8) in Japanese individuals. For \( CDKN2B(AS1) \), rs518394, rs10120806, and rs4977756 could represent all nine SNPs with \( R^2 > 0.8 \) in the Japanese HapMap individuals. Rs10120806 was excluded because a TaqMan probe and primers could not be synthesized. For \( CAV1/CAV2 \), we selected only the rs1052990 with sufficient MAF and statistical power (>0.8) for the Japanese. For \( SIX1/SIX6 \), we selected rs34912345 in addition to the reported one SNP (rs10483727), given multiple reports on the functional activity of rs34912345 to cpRNFLT.\(^{18-20} \) For \( TMCO1 \), the reported two SNPs were monomorphic in the Japanese, and we selected rs12723198, rs10918271, and rs10918281 from 3 LD blocks with \( R^2 > 0.9 \) and MAF > 0 in the Japanese HapMap individuals.

Genomic DNA was prepared from peripheral blood using a DNA extraction kit (QuickGene-610L; Fujifilm, Minato, Tokyo, Japan). For the 756 POAG patients, all the SNPs were genotyped using a TaqMan SNP assay with the ABI PRISM 7700 system (Applied Biosystems, Foster City, CA, USA). Sample call rates of ≤0.9 were excluded. For the 3094 controls in the Nagahama study, genotyping and quality control methods were performed as described.\(^{20} \)

**Statistical Analyses**

For the assessment of genetic associations with cpRNFLT and VFD using case groups, GEE analyses were applied for a both-eyes model incorporating repeated measurements of the paired-eye data.\(^{21-27-29} \) Also, linear regression analyses were performed for a worse-eye model. For the case-control replication analyses, logistic regression analyses were performed assuming multiplicative models for the effect of the minor allele with adjustments for age and sex. A \( P \) value < 0.05 was considered statistically significant for each SNP in the replication analyses.

For both the GEE analyses and linear regression analyses, the additive effect of the per minor allele was assumed with adjustments for age, sex, and axial length at each visit. We evaluated 12 divided sectors along the OCT circle scan, and Bonferroni corrections were applied to the 9 SNPs and 12 sectors. For the SNPs that were significantly \( (P < 4.6 \times 10^{-14}; 0.05/9 \text{ SNPs/12 \text{ sectors}}) \) or marginally \( (P < 4.2 \times 10^{-3}; 0.05/12 \text{ sectors}) \) associated with 12-sectored cpRNFLT, their associations were further evaluated by the analysis of 768 sectors of the cpRNFLT. For the SNPs that were significantly associated \( (P < 4.6 \times 10^{-4}) \) with a cpRNFLT in 768 sectors, their associations with corresponding VFD were analyzed to confirm these regional signals. For this purpose, the associations between these SNP variances and visual field sensitivities of \( 10^\circ \) or \( 24^\circ \) and their upper and lower hemifields were investigated. A \( P \) value < 0.05 was considered statistically significant for these analyses.

We used PLINK\(^{30} \) for the case-control analyses and the R software (http://www.R-project.org/; in the public domain; accessed September 4, 2015) for the other analyses.

**RESULTS**

There were 740 POAG cases and 2723 controls after standard quality controls. The demographic information on these participants is shown in Table 1.

**Case–Control Confirmation Analyses of Association With Glaucoma**

The results of the case-control replication analyses of the glaucoma-susceptible SNPs are shown in Table 2. There were significant associations between the presence of POAG and the SNP genotypes of rs518394 and rs4977756 near
TABLE 1. Demographics of Study Participants

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Cases</th>
<th>Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>740</td>
<td>2723</td>
</tr>
<tr>
<td>Age, y*</td>
<td>70.1±11.9</td>
<td>52.5±14.1</td>
</tr>
<tr>
<td>Sex, male/female</td>
<td>340/400</td>
<td>878/1845</td>
</tr>
<tr>
<td>IOP, mm Hg†</td>
<td>15.97±3.54</td>
<td>NA</td>
</tr>
<tr>
<td>Axial length, mm†</td>
<td>24.10±1.12</td>
<td>23.79±1.03</td>
</tr>
<tr>
<td>Central corneal thickness, μm†</td>
<td>527.2±33.6</td>
<td>NA</td>
</tr>
<tr>
<td>Global cpRNFL, μm†</td>
<td>64.5±14.2</td>
<td>NA</td>
</tr>
<tr>
<td>HFA 10.2 MD value, dB‡</td>
<td>–10.24±7.71</td>
<td>NA</td>
</tr>
<tr>
<td>HFA 24-2/30-2 MD value, dB‡</td>
<td>–8.38±7.63</td>
<td>NA</td>
</tr>
<tr>
<td>Visual acuity, logMAR§</td>
<td>0.048±0.331</td>
<td>NA</td>
</tr>
<tr>
<td>Spherical equivalent refraction, D†</td>
<td>–1.57±2.71</td>
<td>–1.02±2.17</td>
</tr>
<tr>
<td>Follow-up period, y§</td>
<td>7.1±4.2</td>
<td>NA</td>
</tr>
<tr>
<td>Anti-glaucoma ophthalmic medication†</td>
<td>1.9±1.2</td>
<td>NA</td>
</tr>
</tbody>
</table>

NA, not applicable; MD, mean deviation; D, diopter.

* These values are shown in mean ± standard deviation. For cases, ages at the time of study inclusion were adopted.

† These values are shown in mean ± standard deviation. For cases, the values at first visit or the nearest ones were adopted and the both-eye data were averaged, if available.

‡ These values are shown in mean ± standard deviation. All acquired data during follow-up were averaged for each patient to calculate each patient value.

§ These values are shown in mean ± standard deviation. The values of the worse eye at the latest visit were adopted.

CDKN2B(AS1) after adjustments for age and sex distribution. The associations of the SNPs of TMCO1, CAV1/CAV2, and SIX1/SIX6 with POAG were not significant.

Association of Glaucoma-Susceptible Polymorphisms With Regional cpRNFLT

A total of 4864 images of 680 patients (1208 eyes) were studied. The associations between the nine SNPs of the four glaucoma-susceptible genes and the 12-divided cpRNFLT sectors are shown in Tables 3 and 4. In the both-eye model, the rs4977756 and rs518394 SNPs of CDKN2B(AS1) were significantly associated with a regional cpRNFLT (TS [temporal superior] 01; $P = 2.6 \times 10^{-4} < 4.6 \times 10^{-4}$, $\beta = 3.02$; 95% confidence interval [CI]: 1.10–4.64 and TS01; $P = 4.63 \times 10^{-4}$ < 4.68 × 10^{-4}, $\beta = 4.29$; 95% CI: 1.89–6.69, respectively). Two SNPs of SIX1/SIX6 had marginal associations ($P < 4.2 \times 10^{-3}$) with a regional cpRNFLT. In the worse-eye model, the rs10120688 SNP of CDKN2B(AS1) and the two SNPs of SIX1/SIX6 had marginal associations ($P < 4.2 \times 10^{-3}$) with a regional cpRNFLT. The SNPs of TMCO1 and CAV1/CAV2 were not significantly associated with any of the 12-divided cpRNFLT sectors of the two models.

Further assessments of the regional signals were performed using 768 cpRNFLT sectors for these significantly or marginally associated SNPs. The sizes and $P$ values of each SNP genotype association with the 768-sector cpRNFLT are shown in Figures 2 and 3. In the both-eye model, rs4977756 near CDKN2B had significant signal peaks ($P < 4.6 \times 10^{-4}$) in the temporal and nasal regions at 330° to 360°, 0° to 30°, and 150° to 180° (maximum $\beta = 2.92$ [95% CI: 1.55–4.29], $P = 2.9 \times 10^{-7}$ at 351.1°, maximum $\beta = 3.97$ [95% CI: 1.86–6.07], $P = 2.2 \times 10^{-4}$ at 23.4°, and maximum $\beta = 3.50$ [95% CI: 1.55–5.46], $P = 4.2 \times 10^{-4}$ at 166.4°, respectively). The rs518394 SNP near CDKN2B(AS1) had significant signal peaks ($P < 4.6 \times 10^{-4}$) in the temporal region at 330° to 360° and 0° to 30° (maximum $\beta = 3.98$ [95% CI: 1.92–6.04], $P = 1.5 \times 10^{-4}$ at 350.2° and maximum $\beta = 5.31$ [95% CI: 2.35–8.27], $P = 4.4 \times 10^{-4}$ at 22.9°,

TABLE 2. Genotype Counts of the Glaucoma-Related Single Nucleotide Polymorphisms Among Primary Open-Angle Glaucoma Patients and Controls

<table>
<thead>
<tr>
<th>CHR</th>
<th>SNP</th>
<th>Gene</th>
<th>G</th>
<th>T</th>
<th>C</th>
<th>Allele Count</th>
<th>MA</th>
<th>Case</th>
<th>Control</th>
<th>Case</th>
<th>Control</th>
<th>Case</th>
<th>Control</th>
<th>Genotype Count</th>
<th>OR_Nominal</th>
<th>P Value</th>
<th>OR_Adjusted</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1</td>
<td>rs7221099</td>
<td>CDKN2B</td>
<td>137/230/592</td>
<td>57/654/2012</td>
<td>0.104</td>
<td>0.141</td>
<td>0.034</td>
<td>0.67</td>
<td>0.64 (0.55–0.75)</td>
<td>2.3×10−10</td>
<td>0.0018</td>
<td>0.80 (0.71–0.90)</td>
<td>0.020</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>rs10120688</td>
<td>CDKN2B</td>
<td>159/249/502</td>
<td>64/1019/1500</td>
<td>0.235</td>
<td>0.38</td>
<td>0.034</td>
<td>0.67</td>
<td>0.78 (0.68–0.86)</td>
<td>2.4×10−3</td>
<td>0.0108</td>
<td>0.75 (0.65–0.90)</td>
<td>0.29</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>rs977756</td>
<td>SIX1/SIX6</td>
<td>32/239/460</td>
<td>152/1001/1570</td>
<td>0.281</td>
<td>0.250</td>
<td>0.210</td>
<td>0.65</td>
<td>0.91 (0.82–0.99)</td>
<td>0.0021</td>
<td>0.89 (0.78–1.02)</td>
<td>0.35</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>4</td>
<td>rs10483727</td>
<td>SIX1/SIX6</td>
<td>32/245/488</td>
<td>156/992/1576</td>
<td>0.211</td>
<td>0.239</td>
<td>0.190</td>
<td>0.91</td>
<td>0.82 (0.73–0.94)</td>
<td>0.0015</td>
<td>0.81 (0.72–0.90)</td>
<td>0.082</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

CHR, chromosome; HWE, Hardy-Weinberg equilibrium.

† These values are shown in bold.
‡ Logistic regression analysis assuming multiplicative effect of the per minor allele variant, adjusted for age and sex.

** Significant association.
respectively. For the two SNPs of SIX1/SIX6, nonsignificant association signals were observed in the inferior region.

In the worse-eye model, rs10120688 near CDKN2B(AS1) had significant signal peaks \( (P < 4.6 \times 10^{-4}) \) between the temporal to superior region at 60° to 90° (maximum \( f = 10.10 \) [95% CI: 5.12-15.08], \( P = 8.0 \times 10^{-5} \) at 73.6°). The rs33912345 SNP near SIX1/SIX6 had significant signal peaks \( (P < 4.6 \times 10^{-4}) \) in the inferior region at 270° to 300°.

**Association of Glaucoma-Susceptible Polymorphisms With Locations of Visual Field Defects**

Significant \( (P < 4.6 \times 10^{-4}) \) associations of the SNPs near CDKN2B(AS1) and SIX1/SIX6 were confirmed by the severity of VF defects in the corresponding regions. In the better-eye model, 8743 reliable visual field tests of 650 patients (1155 eyes) were included for HFA 24-2/30-2 analyses, whereas 2042 reliable visual field tests of 414 patients (690 eyes) were included for HFA 10-2 analyses. In agreement with the association peaks around 350° and 20° cpRNFLT, a risk allele of rs4977756 near CDKN2B had significant associations to both worse paracentral and paracentral lower hemifield scotomas \( (P < 0.05, \text{ Table } 5) \).

In the worse-eye model using 650 tests of HFA 24-2/30-2 and 414 tests of HFA 10-2, only a risk allele of rs10120688 near CDKN2B(AS1) had a marginal association with the lower hemifield scotoma, and no other significant associations were observed for the other SNPs \( (P > 0.05, \text{ Table } 6) \).

**Discussion**

Our results showed a significant gene-structure relationship between the risk allele of the CDKN2B(AS1) gene and the thinner RNFL in the temporal and superior sectors. Further evaluations by finer sectors of the cpRNFLT and VFD led to our finding that the risk allele of rs4977756 was significantly associated with the worse paracentral and paracentral lower hemifield VFDs.

To date, genetic studies on glaucoma have evaluated not only case-control studies on POAG patients but also various endophenotypes such as the IOP, VCDR, disc area, central corneal thickness, VFD, and cpRNFLT. Among these, the analysis of the cpRNFLT has advantages because of its ability to detect early disease and it also has a linear relationship to the GON, is highly reproducible, and has a wider examined area than the HFA regions. Although the changes in VCDR represent the progression of GON and glaucoma diagnosis, finer analyses using 768-point cpRNFLT would be a better endophenotype for genetic analysis of POAG.

**Table 3.** Associations of Glaucoma-Related Genes and Circumpapillary Retinal Nerve Fiber Layer Thickness Using Repeated Measurements of Both Eyes in 12 Sectors

<table>
<thead>
<tr>
<th>CHR</th>
<th>SNP</th>
<th>Gene</th>
<th>TS 01^*</th>
<th>TS 02^*</th>
<th>TS 03^*</th>
<th>NS 01^*</th>
<th>NS 02^*</th>
<th>NS 03^*</th>
<th>NI 01^*</th>
<th>NI 02^*</th>
<th>NI 03^*</th>
<th>TI 01^*</th>
<th>TI 02^*</th>
<th>TI 03^*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>rs12723198</td>
<td>TMCO1</td>
<td>0.29</td>
<td>0.84</td>
<td>0.98</td>
<td>0.74</td>
<td>0.92</td>
<td>1.00</td>
<td>0.77</td>
<td>0.91</td>
<td>0.77</td>
<td>0.75</td>
<td>0.72</td>
<td>0.81</td>
</tr>
<tr>
<td>1</td>
<td>rs10918271</td>
<td>TMCO1</td>
<td>0.34</td>
<td>0.25</td>
<td>0.43</td>
<td>0.99</td>
<td>0.63</td>
<td>0.50</td>
<td>0.26</td>
<td>0.23</td>
<td>0.52</td>
<td>0.47</td>
<td>0.74</td>
<td>0.86</td>
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<tr>
<td>1</td>
<td>rs10918281</td>
<td>TMCO1</td>
<td>0.67</td>
<td>0.97</td>
<td>0.75</td>
<td>0.73</td>
<td>0.86</td>
<td>0.76</td>
<td>0.99</td>
<td>0.79</td>
<td>0.71</td>
<td>0.81</td>
<td>0.80</td>
<td>0.48</td>
</tr>
<tr>
<td>7</td>
<td>rs1052990</td>
<td>CAV1/C2V2</td>
<td>0.42</td>
<td>0.26</td>
<td>0.16</td>
<td>0.99</td>
<td>0.25</td>
<td>0.26</td>
<td>0.28</td>
<td>0.27</td>
<td>0.62</td>
<td>0.49</td>
<td>0.47</td>
<td>0.65</td>
</tr>
<tr>
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<td>CDKN2B</td>
<td>0.0043</td>
<td>0.11</td>
<td>0.53</td>
<td>0.70</td>
<td>0.74</td>
<td>0.48</td>
<td>0.57</td>
<td>0.62</td>
<td>0.0062</td>
<td>0.14</td>
<td>0.058</td>
<td>0.020</td>
</tr>
<tr>
<td>9</td>
<td>rs33912345</td>
<td>SIX1/SIX6</td>
<td>0.014</td>
<td>0.0068</td>
<td>0.0025</td>
<td>0.14</td>
<td>0.50</td>
<td>0.20</td>
<td>0.90</td>
<td>0.43</td>
<td>0.084</td>
<td>0.37</td>
<td>0.11</td>
<td>0.11</td>
</tr>
</tbody>
</table>

**Table 4.** Associations of Glaucoma-Related Genes and Circumpapillary Retinal Nerve Fiber Layer Thickness of the Worse Eye in 12 Sectors

<table>
<thead>
<tr>
<th>CHR</th>
<th>SNP</th>
<th>Gene</th>
<th>TS 01^*</th>
<th>TS 02^*</th>
<th>TS 03^*</th>
<th>NS 01^*</th>
<th>NS 02^*</th>
<th>NS 03^*</th>
<th>NI 01^*</th>
<th>NI 02^*</th>
<th>NI 03^*</th>
<th>TI 01^*</th>
<th>TI 02^*</th>
<th>TI 03^*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>rs12723198</td>
<td>TMCO1</td>
<td>0.29</td>
<td>0.84</td>
<td>0.98</td>
<td>0.74</td>
<td>0.92</td>
<td>1.00</td>
<td>0.77</td>
<td>0.91</td>
<td>0.77</td>
<td>0.75</td>
<td>0.72</td>
<td>0.81</td>
</tr>
<tr>
<td>1</td>
<td>rs10918271</td>
<td>TMCO1</td>
<td>0.34</td>
<td>0.25</td>
<td>0.43</td>
<td>0.99</td>
<td>0.63</td>
<td>0.50</td>
<td>0.26</td>
<td>0.23</td>
<td>0.52</td>
<td>0.47</td>
<td>0.74</td>
<td>0.86</td>
</tr>
<tr>
<td>1</td>
<td>rs10918281</td>
<td>TMCO1</td>
<td>0.67</td>
<td>0.97</td>
<td>0.75</td>
<td>0.73</td>
<td>0.86</td>
<td>0.76</td>
<td>0.99</td>
<td>0.79</td>
<td>0.71</td>
<td>0.81</td>
<td>0.80</td>
<td>0.48</td>
</tr>
<tr>
<td>7</td>
<td>rs1052990</td>
<td>CAV1/C2V2</td>
<td>0.42</td>
<td>0.26</td>
<td>0.16</td>
<td>0.99</td>
<td>0.25</td>
<td>0.26</td>
<td>0.28</td>
<td>0.27</td>
<td>0.62</td>
<td>0.49</td>
<td>0.47</td>
<td>0.65</td>
</tr>
<tr>
<td>9</td>
<td>rs10120688</td>
<td>CDKN2B</td>
<td>0.0043</td>
<td>0.11</td>
<td>0.53</td>
<td>0.70</td>
<td>0.74</td>
<td>0.48</td>
<td>0.57</td>
<td>0.62</td>
<td>0.0062</td>
<td>0.14</td>
<td>0.058</td>
<td>0.020</td>
</tr>
<tr>
<td>9</td>
<td>rs33912345</td>
<td>SIX1/SIX6</td>
<td>0.014</td>
<td>0.0068</td>
<td>0.0025</td>
<td>0.14</td>
<td>0.50</td>
<td>0.20</td>
<td>0.90</td>
<td>0.43</td>
<td>0.084</td>
<td>0.37</td>
<td>0.11</td>
<td>0.11</td>
</tr>
</tbody>
</table>
Gene–Structure Relationship and the Corresponding Structure–Function Relationship

Using the both-eye model, the risk allele of rs4977756 near CDKN2B was strongly associated with a thinning of the cpRNFLT at around 350° and 20°. According to Hood et al., a thinning in this region should lead to VFDs of the paracentral lower hemifield areas, for the lower and upper hemifield could be separated by around −12° cpRNFL location. Indeed, the risk allele of rs4977756 was significantly associated not only to POAG but also to the severity of paracentral and paracentral lower hemifield VFDs (Table 5). These associations could not be observed when we analyzed paracentral region of the Garway-Heath mapping using HFA 24-2/30-2 (Supplementary Table S1). Thus, it would be of great benefit for such patients with risk alleles of CDKN2B to have their visual fields in the central 10° examined more intensively so as not to decrease the quality of vision. On the contrary, using the worse-eye model, the significant genetic association of rs10120688 near CDKN2B(AS1) with temporal superior cpRNFLT and that of rs33912345 near SIX1/SIX6 with inferior cpRNFLT could not be confirmed by the corresponding lower hemifield VFD and upper hemifield VFD, respectively (Table 6; Supplementary Table S2).

Statistical Approach for Repeated Measurement Data

In general, several analytical methods can be applied for repeated measurement data. However, none of them can effectively solve the problems of the randomness and variability of data, the handling of missing or unselected data, and the correlation structure. In addition, some of these models utilized only individuals with complete ophthalmic data at designated time points. It could be inappropriate if the...
mechanism of missing or unselected data depended on the disease-related factors. To deal with as many of these analytical problems as possible, we used GEE analysis for a both-eye model.21 Because POAG is a markedly asymmetric disease but the contribution of genetic factors is equal between the two eyes of a patient, analyzing both eyes would lead to a more accurate estimate that excludes various unilateral effects on environment, lifestyle, dominant eye, and traumatic history of the patients. Therefore, we adopted the GEE model as well as the worse-eye model in this study.

Case–Control Replication Study
In the case–control studies, we observed negative associations of the TMCO1 and CAV1/CAV2 genes with POAG in Japanese individuals. In fact, the associations of CAV1/CAV2 to POAG were contradictory in the following replication studies on Japanese patients with POAG.46,47 For rs1052990 near CAV1/CAV2 and the three SNPs near TMCO1, a type 1 error of 0.05, power of 0.8, and sample/control number of 711/2723, ORs of 1.313 and 1.338 are needed, respectively, to be able to reject the null hypothesis. These ORs are comparable to those in previous reports.5,6,46 Thus, the association of CAV1/CAV2 and TMCO1 to glaucoma is less significant in the Japanese.

There are several limitations to this study. First, earlier studies48,49 showed that the cpRNFLT was inversely correlated with the axial length. In addition, eyes with longer axial length could be affected by the axial length–related magnification of the OCT images.50 Thus, we excluded highly myopic eyes and applied statistical adjustment by axial length. Second, in the analyses of cpRNFLT, we included case-only subjects without healthy controls and preperimetric glaucoma patients. If we had included a wider range of disease severity, glaucoma-susceptible SNPs would inevitably be detrimental to the glaucoma-susceptible cpRNFL regions. Thus, the case-only approach would be appropriate. Third, cpRNFLT of the

FIGURE 3. Effect of size, direction, and P values of the single nucleotide polymorphism (SNP) genotypes to the 768-point circumpapillary retinal nerve fiber layer thicknesses (cpRNFLT) along with the optical coherence tomographic circle scans in the worse-eye model. First line shows the effect sizes and 95% confidence intervals based on minor alleles shown in Table 2. The second line shows the P values plotted in –log10 scale for each SNP. A red line shows P value of 1 × 10⁻³ (< 0.05 / 9 SNPs / 12 sectors = 4.6 × 10⁻⁴). (A) Rs10120688 near CDKN2B(AS1) has significant associations with temporal superior cpRNFLT at 60° to 90°. (B) Rs33912345 near SIX1/SIX6 has significant associations with inferior cpRNFLT at 270° to 300°. (C) Rs10483727 near SIX1/SIX6 have nonsignificant associations with inferior cpRNFLT at 270° to 300°.
### Table 5. Association Results Between Single Nucleotide Polymorphism Genotypes and Visual Field Sensitivity in the Humphrey Field Analyzer 24-2 and 10-2 Plot Using Repeated Measurements of the Both Eyes

<table>
<thead>
<tr>
<th>CHR</th>
<th>SNP</th>
<th>Gene</th>
<th>HFA 24-2, 52 Points</th>
<th>Hemifield 26 Points</th>
<th>HFA 10-2, 68 Points</th>
<th>Paracentral Hemifield 34 Points</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>β Adjusted (95% CI)</td>
<td>P Adjusted†</td>
<td>β Adjusted (95% CI)</td>
<td>P Adjusted†</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Upper</td>
<td>Lower</td>
<td>Upper</td>
<td>Lower</td>
</tr>
<tr>
<td>9</td>
<td>rs518394</td>
<td>CDKN2B-AS1</td>
<td>0.02 (0.00 to 0.07)</td>
<td>0.03 (0.03 to 0.07)</td>
<td>0.02 (0.00 to 0.07)</td>
<td>0.03 (0.02 to 0.07)</td>
</tr>
<tr>
<td>9</td>
<td>rs4977756</td>
<td>CDKN2B</td>
<td>0.01 (0.00 to 0.04)</td>
<td>0.01 (0.03 to 0.05)</td>
<td>0.01 (0.00 to 0.04)</td>
<td>0.02 (0.02 to 0.05)</td>
</tr>
</tbody>
</table>

CHR, chromosome.

* Effect size on the mean values of 52 or 68 points in HFA 24-2/30-2 or 10-2 total deviation plot. The analyses were performed in 1/Lambert scales.

† Results from generalized estimation equation models, which account for all acquired data of the patients assuming additive effect of the per minor allele variant, adjusted for age at each visit, sex, and axial length. Significant (P < 0.05) associations are shown in bold.

‡ Effect size on the mean values of paracentral hemifield 26 or 34 points in HFA 24-2/30-2 or 10-2 total deviation plot. The analyses were performed in 1/Lambert scales.

---

### Table 6. Association Results Between Single Nucleotide Polymorphism Genotypes and Visual Field Sensitivity in the Humphrey Field Analyzer 24-2 and 10-2 Plot of the Worse Eye

<table>
<thead>
<tr>
<th>CHR</th>
<th>SNP</th>
<th>Gene</th>
<th>HFA 24-2, 52 Points</th>
<th>Hemifield 26 Points</th>
<th>HFA 10-2, 68 Points</th>
<th>Paracentral Hemifield 34 Points</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
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<td></td>
<td>β Adjusted (95% CI)</td>
<td>P Adjusted†</td>
<td>β Adjusted (95% CI)</td>
<td>P Adjusted†</td>
</tr>
<tr>
<td></td>
<td></td>
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<td>Upper</td>
<td>Lower</td>
<td>Upper</td>
<td>Lower</td>
</tr>
<tr>
<td>9</td>
<td>rs10120688</td>
<td>CDKN2B-AS1</td>
<td>0.05 (0.00 to 0.09)</td>
<td>0.06 (0.00 to 0.11)</td>
<td>0.04 (0.03 to 0.08)</td>
<td>0.00 (0.04 to 0.05)</td>
</tr>
<tr>
<td>9</td>
<td>rs33912345</td>
<td>SIX1/SIX6</td>
<td>0.02 (0.00 to 0.07)</td>
<td>0.03 (0.02 to 0.07)</td>
<td>0.01 (0.00 to 0.07)</td>
<td>0.02 (0.03 to 0.08)</td>
</tr>
</tbody>
</table>

CHR, chromosome.

* Effect size on the mean values of 52 or 68 points in HFA 24-2/30-2 or 10-2 total deviation plot. The analyses were performed in 1/Lambert scales.

† Results from generalized estimation equation models, which account for all acquired data of the patients assuming additive effect of the per minor allele variant, adjusted for age at each visit, sex, and axial length. Significant (P < 0.05) associations are shown in bold.

‡ Effect size on the mean values of paracentral hemifield 26 or 34 points in HFA 24-2/30-2 or 10-2 total deviation plot. The analyses were performed in 1/Lambert scales.
Associations Between cpRNFL Thickness and Glaucoma Genes

severely advanced glaucoma patients should be affected by “the floor effect.” That is, glaucoma worsening did not correlate with cpRNFL change in advanced cases. To overcome this limitation, we further confirmed the genetic associations with cpRNFL by those with VFD, and only consistent results have been reported. In addition, the global cpRNFL of the worse eye of the patient was normally distributed (P = 0.28, Shapiro-Wilk test). Fourth, several glaucoma-susceptible polymorphisms of CDKN2B(AS1) showed different associations with cpRNFL of the temporal region and the temporal superior region. Although we highlighted only rs4977756 showing significant genetic associations with corresponding VFD regions, the associations of rs10120688 with corresponding VFD clusters were marginal. Thus, further studies should be done to disprove these multifaceted gene–structure relationships.

In conclusion, we showed regional associations of CDKN2B(AS1) to the temporal cpRNFLT at around 350° and 20° that could be differentiated by analyzing the 768 cpRNFL thicknesses along the OCT circle scan, using large numbers of case-only POAG patients. These cpRNFL regional associations of glaucoma-related genes led to the finding that the CDKN2B gene was associated with paracentral and paracentral lower hemisphere scotomas. The cpRNFLT with such fineness would be a better endophenotype for genetic analysis of POAG, and it enabled us to determine the effects of glaucoma-related genes on the region-specific RNFLT.

Acknowledgments

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Associations Between cpRNFL Thickness and Glaucoma Genes


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APPENDIX

The Nagahama Study Group

The following investigators were core members of the Nagahama Study Group: Takeo Nakayama (Department of Health Informatics, Kyoto University School of Public Health, Kyoto, Japan); Akihiro Sekine (Center for Genomic Medicine, Graduate School of Medicine, Kyoto University, Kyoto, Japan); Shinji Kosugi (Department of Medical Ethics, Kyoto University Graduate School of Medicine, Kyoto University, Kyoto, Japan); Ryo Yamada (Center for Genomic Medicine, Graduate School of Medicine, Kyoto University, Kyoto, Japan); and Yasuharu Tabara (Center for Genomic Medicine, Graduate School of Medicine, Kyoto University, Kyoto, Japan).