A Comparison of En Face Optical Coherence Tomography and Fundus Autofluorescence in Stargardt Disease

Vivienne C. Greenstein, 1 Jason Nunez, 2 Winston Lee, 1 Kaspar Schuerch, 1 Brad Fortune, 3 Stephen H. Tsang, 1,4 Rando Allikmets, 1,4 Janet R. Sparrow, 1,4 and Donald C. Hood 1,2

1Department of Ophthalmology, Columbia University, New York, New York, United States
2Department of Psychology, Columbia University, New York, New York, United States
3Devers Eye Institute, Portland, Oregon, United States
4Department of Pathology & Cell Biology, Columbia University, New York, New York, United States

Correspondence: Vivienne C. Greenstein, Department of Ophthalmology, Columbia University, 160 Fort Washington Avenue, Room 508, New York, NY 10032, USA; vcg17@columbia.edu.

Submitted: July 1, 2017
Accepted: September 8, 2017

RESULTS. For all eyes, RPE atrophy was visualized as a central hyperreflective area on the subRPE slab, and IS/OS junction loss as an abnormal reflective area on the IS/OS slab; the latter was significantly larger (P = 0.04). There was good agreement between the hyperreflective area on the subRPE slab image and hypoSW-AF area, and between the abnormal reflective area on the IS/OS slab and hypo-hyperSW-AF area; the hypoNIR-AF area indicated that the hyperreflective area on the subRPE slab underestimated RPE atrophy. The spatial locations of hyperreflective flecks on the en face preRPE(OS) slab image corresponded to those on the SW-AF images.

CONCLUSIONS. Wide-field en face OCT imaging has the potential to be a clinically useful tool for the management of STGD1.

Keywords: wide-field en face optical coherence tomography, Stargardt disease, fundus autofluorescence

Recessive Stargardt disease (STGD1), a form of juvenile macular degeneration, is caused by more than 1000 mutations in the photoreceptor-specific ABCA4 gene, which encodes the ATP-binding cassette transporter in photoreceptors. 1,2 The transporter facilitates the clearance of all-trans-retinaldehyde, 3 which is generated by photon-mediated cis-trans isomerization of retinaldehyde in the photoreceptor cell. When the transport activity is reduced or absent, the result is accumulation of lipofuscin in RPE cells. 4 The transport activity is accompanied by a distribution of yellowish white deposits or “flecks” visible at the posterior pole and sometimes in the mid-periphery. 5 Initially the flecks appear to be well-defined, but with disease progression confluence increases across the posterior pole and they resorb leaving residual atrophy. In addition, their spatial distribution changes over time; typically they spread in a centrifugal direction from the fovea, 6,7 but other patterns have been found in individual patients. 5

Several noninvasive imaging modalities are used for the diagnosis and monitoring of disease progression in STGD1. These include short wavelength fundus autofluorescence (SW-AF), near infrared autofluorescence (NIR-AF), 8 and spectral domain optical coherence tomography (SD-OCT). SW-AF imaging reveals areas of decreased autofluorescence or hypofluorescence (hypoSW-AF), and areas of hyperautofluorescence (hyperSW-AF) in the form of flecks and/or rings. With SD-OCT it has been shown that the area of abnormal SW-AF (total hypoSW-AF and hyperSW-AF) in the central macula is associated with loss of the ellipsoid zone band (aka inner/outer segment [IS/OS] junction) and loss of the RPE as revealed by NIR-AF imaging. 9-11 In one study, a high correlation between photoreceptor loss and the extent of abnormal SW-AF was found. 12

However, the exact sequence of retinal degeneration occurring with disease progression remains controversial.
Depending on the imaging modality, it has been suggested that photoreceptor loss occurs before RPE loss,20,21 that RPE cell loss and/or dysfunction occurs and then there is secondary photoreceptor cell loss,22,23 and that changes in the photoreceptor layer may occur simultaneously with the development of abnormalities in the RPE layer.20 In addition, the intraretinal receptor layer may occur simultaneously with the development of abnormalities in the RPE layer.20–22 However, in NIR-AF images, flecks are the RPE, it has been assumed that they are to be found at the position of the hyperSW-AF flecks, which often are observed on SW-AF also is of interest. Since the autofluorescent appearance of flecks usually is associated with lipofuscin in the RPE, it has been assumed that they are to be found at the level of the RPE.20–22 However, in NIR-AF images, flecks are dark due to loss of RPE, while in horizontal SD-OCT images they correspond to hyperreflective deposits that are anterior to RPE/Bruch’s membrane and traverse photoreceptor-attributable OCT layers.12,24 There now is abundant evidence in human disease and mouse models that degenerating photoreceptor cells generate bisretinoid at elevated levels and it is reasonable to suggest that the SW-AF of flecks originates in degenerating photoreceptor cells.

We used an en face imaging approach to gain further insight into the morphologic changes that accompany macular atrophy, and corroborated our understanding of the intraretinal location of flecks in patients with STGD1. Advances in OCT technology, such as increased speed of acquisition of images, improved resolution, and improvements in software and data processing, have enabled rapid and powerful visualization of en face OCT projection images. The en face visualization approach provides a coronal view of a particular layer within the central retina by isolating its reflections from a specified depth. Thus, it aids in the precise localization and measurement of lesions within specific retinal layers. It also provides the ability to register the resulting en face images to other fundus imaging modalities, such as fundus autofluorescence (FAF), using retinal vessels and other landmarks (e.g., see prior studies24,25). In this study, we compared the morphologic changes visualized on en face images derived from wide-field swept-source (ss) OCT scans qualitatively and quantitatively to hypo- and hyperautofluorescent (hypoAF, hyperAF) areas detected with SW-AF.

### METHODS

#### Subjects

We studied 16 patients (16 eyes; 9 male and 7 female patients; mean age, 38.3 ± 17.7 years; range, 11–70 years), with at least one (expected) disease-causing mutation in the *ABCA4* gene. Demographic, genetic, and selected clinical data are presented in Table 1. All patients had a complete ocular examination, including best-corrected visual acuity with subjective refraction. Eyes were excluded from the study if there was evidence of significant ocular media opacities, refractive errors greater than ±6 diopter (D) sphere or ±2 D cylinder, elevated IOP > 21 mm Hg, and a history or diagnosis of any other significant ocular comorbidities. The eye with the higher image quality was used for analysis.

<table>
<thead>
<tr>
<th>Patient</th>
<th>Sex</th>
<th>Age</th>
<th>Decade</th>
<th>Race/Ethnicity</th>
<th>Study Eye</th>
<th>Snellen</th>
<th>logMAR</th>
<th>fERG Group†</th>
<th>Clinical Stage‡</th>
<th>ABCA4 Mutations</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>F</td>
<td>56</td>
<td>5th</td>
<td>White</td>
<td>OS</td>
<td>20/40</td>
<td>0.30</td>
<td>1</td>
<td>1</td>
<td>p.R2077W</td>
</tr>
<tr>
<td>2</td>
<td>M</td>
<td>45</td>
<td>4th</td>
<td>Indian</td>
<td>OD</td>
<td>20/25</td>
<td>0.01</td>
<td>1</td>
<td>1</td>
<td>p.R511H</td>
</tr>
<tr>
<td>3</td>
<td>F</td>
<td>16</td>
<td>1st</td>
<td>White</td>
<td>OS</td>
<td>20/125</td>
<td>0.80</td>
<td>2</td>
<td>1</td>
<td>p.R1108C</td>
</tr>
<tr>
<td>4</td>
<td>F</td>
<td>56</td>
<td>4th</td>
<td>Indian</td>
<td>OD</td>
<td>20/25</td>
<td>0.01</td>
<td>N/A</td>
<td>3</td>
<td>p.P1486L</td>
</tr>
<tr>
<td>5</td>
<td>M</td>
<td>11</td>
<td>1st</td>
<td>White</td>
<td>OS</td>
<td>20/100</td>
<td>0.70</td>
<td>1</td>
<td>1</td>
<td>p.[L541P,A1038V]</td>
</tr>
<tr>
<td>6</td>
<td>M</td>
<td>16</td>
<td>1st</td>
<td>Indian</td>
<td>OS</td>
<td>20/60</td>
<td>0.48</td>
<td>2</td>
<td>2</td>
<td>p.[L541P,A1038V]</td>
</tr>
<tr>
<td>7</td>
<td>F</td>
<td>51</td>
<td>5th</td>
<td>White</td>
<td>OD</td>
<td>20/25</td>
<td>0.01</td>
<td>1</td>
<td>1</td>
<td>p.[L541P,A1038V]</td>
</tr>
<tr>
<td>8</td>
<td>M</td>
<td>25</td>
<td>2nd</td>
<td>White</td>
<td>OS</td>
<td>20/70</td>
<td>0.54</td>
<td>1</td>
<td>1</td>
<td>p.W15*</td>
</tr>
<tr>
<td>9</td>
<td>M</td>
<td>38</td>
<td>3rd</td>
<td>White</td>
<td>OS</td>
<td>20/100</td>
<td>0.70</td>
<td>N/A</td>
<td>1</td>
<td>p.G1961E</td>
</tr>
<tr>
<td>10</td>
<td>M</td>
<td>50</td>
<td>4th</td>
<td>African-American</td>
<td>OS</td>
<td>20/400</td>
<td>1.50</td>
<td>1</td>
<td>1</td>
<td>p.T1526M</td>
</tr>
<tr>
<td>11</td>
<td>M</td>
<td>70</td>
<td>5th</td>
<td>White</td>
<td>OD</td>
<td>20/80</td>
<td>0.60</td>
<td>1</td>
<td>1</td>
<td>p.W663*</td>
</tr>
<tr>
<td>12</td>
<td>F</td>
<td>47</td>
<td>3rd</td>
<td>Indian</td>
<td>OD</td>
<td>20/100</td>
<td>0.70</td>
<td>1</td>
<td>1</td>
<td>c.2069delG</td>
</tr>
<tr>
<td>13</td>
<td>M</td>
<td>30</td>
<td>3rd</td>
<td>Indian</td>
<td>OS</td>
<td>20/70</td>
<td>0.54</td>
<td>1</td>
<td>1</td>
<td>p.R653C</td>
</tr>
<tr>
<td>14</td>
<td>M</td>
<td>54</td>
<td>3rd</td>
<td>Indian</td>
<td>OD</td>
<td>20/400</td>
<td>1.50</td>
<td>1</td>
<td>1</td>
<td>p.V1706*</td>
</tr>
<tr>
<td>15</td>
<td>F</td>
<td>26</td>
<td>2nd</td>
<td>White</td>
<td>OD</td>
<td>20/100</td>
<td>0.70</td>
<td>1</td>
<td>1</td>
<td>p.G1961E</td>
</tr>
<tr>
<td>16</td>
<td>F</td>
<td>22</td>
<td>2nd</td>
<td>White</td>
<td>OD</td>
<td>20/100</td>
<td>0.70</td>
<td>1</td>
<td>1</td>
<td>p.G1961E</td>
</tr>
</tbody>
</table>

F, female; M, male; OS, left eye; OD, right eye; fERG, full-field electroretinogram; N/A, not available.

* Denotes nonsense mutation.
† Lois et al.27
‡ Fishman GA.28
§ Siblings.

Full-field scotopic and photopic electroretinograms (ERGs) were obtained from both eyes of 14 patients according to the International Society for Clinical Electrophysiology of Vision (ISCEV) standards.26 Based on the full-field ERG results, patients were classified into group 1 if the scotopic and photopic ERG amplitudes were within normal limits (12 patients, 12 eyes), group 2 if the photopic ERG amplitudes were decreased (patients P3 and P6), or group 3 if the scotopic and photopic ERG amplitudes were decreased.27 In addition the disease was classified into one of four clinical stages or phenotypes based on those described by Fishman.28 A total of 12 patients had stage 1 (characterized by pigmentary changes in the macula ranging from nonspecific pigment molting to the presence of small atrophic-appearing foveal lesions with localized parfoveal or perifoveal flecks), three stage 2 (presence of numerous yellowish-white flecks throughout the posterior pole) and one stage 3 (extensive macular atrophy with diffuse flecks throughout the foveus) disease (Table 1).

The study was performed under protocol AAA9906 with the approval of the institutional review board of Columbia University. All study procedures complied with the Health
Insurance Portability and Accountability Act of 1996. All patients were enrolled in accordance with the tenets set out in the Declaration of Helsinki. Informed consent was obtained before enrollment.

**Genetic Analyses**

All patients were screened for $ABCA4$ variants by complete sequencing of all coding and intron/exon boundaries of the gene by either Sanger or next-generation sequencing (NGS) as described previously, or with the Illumina TruSeq Custom Amplicon protocol (Illumina, San Diego, CA, USA), followed by sequencing on the Illumina MiSeq platform. The NGS reads were analyzed and compared to the reference genome GRCh37/hg19, using the variant discovery software NextGENe (SoftGenetics LLC, State College, PA, USA). All detected possibly disease-associated variants were confirmed by Sanger sequencing and analyzed with Alamut software (available in the public domain at http://www.interactive-biosoftware.com). Segregation of the variants with the disease was analyzed if family members were available. The allele frequencies of all variants were compared to the Exome Variant Server (EVS) dataset, NHLBI Exome Sequencing Project, Seattle, WA, USA (available in the public domain at http://snp.gs.washington.edu/EVS/; accessed March 2014). Genetic data are presented in Table 1. Two expected disease-causing $ABCA4$ variants were detected in 11 patients and one mutation in the remaining five patients. Patient genotypes consisted mostly of known missense variants including the c.5882G>A (p.G1961E), harbored by six patients (P8, P9, P13, P14, P15, and P16), in addition to four stop-gain mutations and two small deletions, c.2069delG and c.6148-698_c.6670del - a 4770 base pair (bp) deletion.

**Imaging**

Following pupil dilation, SW-AF (488 nm excitation) 30° and 55° images were acquired with the Spectralis HRA+OCT (Heidelberg Engineering, Heidelberg, Germany) after a 20-second bleach of the photopigments in AF mode. Care was taken to obtain high-quality images with maximum field uniformity. In addition, a 9-mm high-resolution horizontal line scan through the fovea was obtained with the Spectralis. The SD-OCT scans were registered automatically to a simultaneous-ly acquired near-infrared reflectance (NIR-R) fundus image. In addition NIR-AF (787 nm excitation) images were acquired from 6 of the 16 eyes with the Heidelberg Retina Angiograph 2 scanning laser ophthalmoscope (HRA2-cSL0, Heidelberg Engineering) using the indocyanine-green angiography mode (without injection of dye) after focus adjustment in infrared reflectance mode.

Wide-field (12 × 9 mm) cube scans consisting of 256 B-scans, each with 512 A-scans, were obtained from all 16 eyes with ssOCT (DRI-OCT; Topcon, Inc., Tokyo, Japan). Following manual correction of the automated segmentation of the IS/OS junction, OS/RPE, and RPE/BM boundaries, special purpose software (ATL 3D-Suite; Fortune B, et al. *IOVS* 2014;55:ARVO E-abstract 2644) was used to generate en face slab images based on the average reflectance intensity for slabs of constant thickness. To obtain optimal visualization of the morphologic changes, we chose the following en face slabs: A subRPE slab with a thickness of 208 μm below the RPE/BM boundary; an IS/OS slab positioned on the IS/OS junction with a thickness of 13 μm; and a preRPE(OS) slab positioned just above the proximal RPE border and protruding into the IS/OS region with a thickness ranging from 13 μm. Examples of the three slabs together with their en face images for a healthy control are shown in Figures 1A to 1C. The thicknesses of the slabs were chosen based on a pilot study and on those used by Sodi et al. The subRPE slab image was used to measure the central RPE atrophy (i.e., on the subRPE slab image the atrophic RPE area was visualized indirectly as a “hyperreflective area”, the increased OCT signal resulted from the loss of the RPE and choriocapillaris); the IS/OS slab image was used to measure the area of IS/OS junction loss; and the preRPE(OS) slab image was used to evaluate the spatial and intraretinal location of flecks. The areas of RPE atrophy and IS/OS junction loss were measured in mm² by two independent observers using ImageJ64 (National Institutes of Health [NIH], Bethesda, MD, USA; available in the public domain at imagej.nih.gov/ij/). The quality of some of the en face IS/OS slab images affected the clarity of the boundaries of the area of IS/OS junction loss. For these cases, the locations chosen to mark the boundary were checked by comparing them to the locations of IS/OS junction loss on the corresponding individual B-scans. The areas of RPE atrophy and IS/OS junction loss then were compared respectively to the area of hypoAF (i.e., absent AF on the SW-AF image and central darkened area on the NIR-AF image) and

![Figure 1](http://tvst.arvojournals.org/ on 01/21/2018)
to the area of abnormal AF (i.e., the hypo- and surrounding hyperAF areas on the SW-AF image). The latter also were measured using ImageJ64. Lastly the spatial and intraretinal locations of flecks observed on the preRPE(OS) slab image were compared to those on the SW-AF image. All en face, SW-AF, and NIR-AF images were aligned to each other using i2kRetina software (DualAlign LLC, Clifton Park, NY, USA) and Adobe Photoshop CS5 (Adobe, Mountain View, CA, USA).

Statistical Analyses
Statistical analysis was performed using PRISM 5 (GraphPad Software, Inc., La Jolla, CA, USA). Bland-Altman plots were generated to assess the agreement between the en face and SW-AF measurements. The statistical significance of the differences between the subRPE and IS/OS areas, and between the hypoAF and total abnormal AF areas was tested with paired-samples t-tests using SPSS Statistics 20.0 for Mac (SPSS IBM, Chicago, IL, USA). In addition, to assess the agreement between the measurements made by the two observers, interclass correlation coefficients (ICC) were calculated in SPSS.

RESULTS
Changes on SW-AF
All 16 eyes exhibited changes on SW-AF. The SW-AF images for 11 of the 16 eyes showed a central area of hypoAF (yellow arrow) surrounded by a well-defined ring of hyperAF (red arrow). The central hypoAF area on the SW-AF image for P8 and the central area of abnormal AF are outlined in the insets. (B) The central area is surrounded by a mottled pattern of flecks (red arrow). (C, D) The fovea is encircled by hyper- and hypoAF flecks in the perifovea of varying size and shape. (E) Mottling on the central fovea. (F) Widespread hypo- and hyperAF changes.

Table 2. Summary of SW-AF and En Face OCT Measurements

<table>
<thead>
<tr>
<th>Short Wavelength-Autofluorescence (SW-AF)</th>
<th>SubRPE Slab</th>
<th>IS/OS Slab</th>
</tr>
</thead>
<tbody>
<tr>
<td>HypoAF</td>
<td>Abnormal AF</td>
<td>Central Hyperreflective Area</td>
</tr>
<tr>
<td>n</td>
<td>11</td>
<td>11</td>
</tr>
<tr>
<td>Mean</td>
<td>4.66 mm²</td>
<td>14.37 mm²</td>
</tr>
<tr>
<td>SD</td>
<td>4.57 mm²</td>
<td>13.18 mm²</td>
</tr>
</tbody>
</table>

* The abnormal SW-AF area was significantly larger than the hypoAF area ($P = 0.01$).
† The IS/OS area was significantly larger than the subRPE area ($P = 0.04$).
**Figure 3.** NIR and SW-AF images for P11 and P14.

**Figure 4.** SubRPE, IS/OS en face, and SW-AF images for one patient, P4. The yellow arrows on the subRPE, IS/OS, and SW-AF images indicate foveal sparing. The subRPE slab image shows a central hyperreflective area. The corresponding en face IS/OS slab image shows a central relatively hyporeflective area surrounded by a hyperreflective border that appears to be larger than the area on the subRPE slab image. The SW-AF image shows a large central hypoAF area. The lower panels show horizontal ssOCT B-scans and respective subRPE slab and IS/OS slab.
and mottling on the foveal center with no other visible areas of hypo-or hyperAF was observed in another eye (Fig. 2E). The areas of abnormal AF (i.e., the hypo- and surrounding hyperAF areas) and the areas of central hypoAF or absent AF were measured (see examples in Figs. 2A, 2B inserts). Unsurprisingly, for the 11 eyes with a central hypoAF area, the abnormal AF area was significantly larger (mean = 14.37 mm², SD = 13.18 mm²) than the hypoAF area (mean = 4.66 mm², SD = 4.57 mm²; P = 0.01; Table 2). For the six eyes with NIR-AF, the darkened areas in the NIR-AF images, indicating a reduction or absence of the NIR-AF signal, exhibited good correspondence to the areas of abnormal AF in the SW-AF images (see examples P11 and P14, Fig. 3).

**OCT En Face Slabs**

The subRPE slab images for the 11 eyes with central hypoAF on SW-AF showed a central hyperreflective area (see Figs. 4, 5A-D). The hyperreflectivity of this zone is attributable to scleral reflectance due to penetration of light into the choroid due to loss of the RPE and atrophy of the choriocapillaris as visualized in the ssOCT B-scan (Fig. 4, bottom left). As can be seen in Figures 4 and 5, the hyperreflective areas were similar in size and shape to the central hypoAF areas on SW-AF. For the subgroup of eyes with NIR-AF, these hyperreflective areas were smaller than the hypoNIR-AF areas.

Foveal sparing, present in four eyes (P1, P2, P4, and P7), also was an identifiable feature on the en face image. It was discernable as a hyporeflective area in the fovea, and as a relatively hyperAF area in the SW-AF image (Fig. 4, yellow arrows). Of interest, the subRPE slab image for the eye with widespread hypo- and hyperAF changes (P6, Fig. 2F) showed a well-defined central hyperreflective area surrounded by hypo- and hyperreflective patterning (Fig. 5A), while the eye with foveal mottling on SW-AF (P5, Fig. 2E) exhibited an ill-defined central hyporeflective area on the subRPE slab image.
The morphologic changes on en face images derived from wide-field ssOCT scans were compared qualitatively and quantitatively to changes visible on SW-AF. In agreement with previous reports, a central hyperreflective area attributable to RPE atrophy was observed on the en face subRPE slab image (Figs. 4, 5). The area was similar in size and shape to the hypoAF area in the central macula in the SW-AF image. Consistent with observations by Sodi et al., measurements of the hyperreflective area on the en face subRPE slab image (i.e., the area of RPE atrophy) revealed that this lesion area was significantly smaller than the area of abnormal reflectance on the IS/OS slab image (i.e., the area related to IS/OS junction loss). However, it also was smaller than the area of RPE cell loss identified by NIR-AF.

Our findings provided some insight into the possible sequence of outer retinal degenerative changes in STGD1. First, we assume that IS/OS junction loss is correlated with photoreceptor loss and/or dysfunction; therefore, hypo–SW-AF or absent SW-AF in STGD1 represents RPE loss and/or atrophy; that hyper–SW-AF can be attributed to accelerated bisretinoid production from degenerating photoreceptor cells; and that it is associated with RPE cells being absent or with thinning and dysfunctional RPE. Second, our SW-AF results indicated that the area of abnormal AF was significantly larger than the hypo- or absent AF area, and that it included the area of IS/OS junction loss. However, these observations cannot be interpreted as evidence that photoreceptor degeneration precedes RPE atrophy in STGD1. Additionally, the tested eye of P5 at the initial visit showed IS/OS junction loss in the fovea on ssOCT and SD-OCT, and central mottling in the SW-AF image that
would be indicative of RPE atrophy (see Figs. 2D, 5D). This interpretation raises questions about the results of previous studies comparing NIR- and SW-AF in patients with STGD1. In these studies, the zone of decreased NIR-AF was reported to be larger than the zone of reduced or abnormal SW-AF and more closely related to the extent of the IS/OS junction loss seen on SD-OCT.13,14 As the main source of NIR-AF is melanin in the RPE and choroid, it was suggested that the results were consistent with RPE cell loss preceding photoreceptor cell degeneration.13,14 In the current study, NIR-AF images were obtained from six of the 16 patients and in agreement with previous studies the central area of absent or hypoAF was larger on NIR than SW-AF.13,14 In addition, the lesion area, indicated by darkening in the NIR-AF image, was larger than the en face subRPE area. When the surrounding clusters of hypoAF, that is, patchy hypoAF areas on NIR-AF, were included in the measurement, the total area was greater than either the en face IS/OS or abnormal SW-AF areas (see Fig. 3). Since the NIR-AF signal is produced, for the most part, from RPE melanin, absence of NIR-AF is indicative of loss of RPE cells. Residual levels of NIR-AF could be accounted for readily by melanin-containing RPE debris from the degenerating cells. If the latter is assumed, the combined NIR-, SW-AF and en face results suggested that photoreceptor cell degeneration has occurred in the presence of dead and/or dying dysfunctional RPE cells.

We also were interested in determining whether the wide-field en face technique could be used to corroborate our previous findings related to the intraretinal location of flecks in patients with STGD1.7 These retinal flecks are an important clinical feature of STGD1. We found that the en face preRPE(OS) slab image provided the best means for detecting flecks. They were evident as small hyperreflective structures either surrounding the central hyporeflective area or widely distributed throughout the image. A hyporeflective border surrounded some of the structures. They corresponded spatially to hyperAF flecks identified on SW-AF images (see examples, Figs. 7A, 7C, 7D) and to darkened foci in NIR-AF images as has been described previously.7 With regard to their intraretinal location, it had been assumed that the AF of flecks in SW-AF images originated from RPE cells since SW-AF is associated with lipofuscin in the RPE.20–22 However, horizontal ss- and SD-OCT B-scans through the fovea (Figs. 7A–D, lower), revealed that, in some cases, the flecks extended anteriorly through photoreceptor-attributable bands and interrupted the IS/OS junction band, and even the band attributable to the outer limiting membrane (Fig. 7D). These cases are in agreement with those of Sparrow et al.,7 who reported that flecks identified in SW-AF images corresponded to hyperreflective deposits traversing photoreceptor-attributable bands in SD-OCT scans. The latter lesions are interpreted as being indicative of degenerating photoreceptor cells.

Our results using the wide-field en face approach have implications for the clinician. They suggested that en face wide-field ssOCT has the potential to be a clinically useful tool in the management of patients with STGD1. In agreement with recent studies, we have shown that the subRPE slab enhances visualization of RPE atrophy.18,35 We also have shown that the IS/OS slab provides the best means for visualizing IS/OS junction loss, and the preRPE(OS) slab for flecks. In addition, when we compared the en face sub-RPE measurements to the central hypoAF area measurements on SW-AF, and the en face...
IS/OS measurements to the abnormal AF measurements, we found good agreement. The results of the Bland-Altman plots suggested that the en face wide-field ssOCT technique can be used as a complementary test to SW-AF, the current clinical gold standard, for measuring and monitoring changes in RPE atrophy. Moreover, the en face approach we used has the added advantage that the clinician can visualize and measure the area of IS/OS junction loss, a marker of photoreceptor loss, inspect the cross-sectional scans to obtain information regarding the status of the RPE and photoreceptor layers, and determine the intraretinal location of flecks. The use of the wide-field ssOCT technique can serve as a complimentary test to SW-AF for detecting and monitoring changes at various stages of the disease whether SW-AF changes present as moderately defined mottling as in Figure 2E, or are widespread as in Figure 2F. In both cases, we were able to observe, and measure, a central hyperreflective area indicating RPE atrophy on the en face subRPE slab image.

One of the limitations of the en face approach is that it is dependent on reliable segmentation of the IS/OS junction band and RPE/BM boundary. We found that we had to correct some of the automated segmentation of these boundaries manually, particularly in the region of the lesion. In addition, the quality of some of the en face IS/OS slab images affected the clarity of the boundaries of IS/OS junction loss. For these cases, we checked the accuracy of our en face IS/OS area measurements by comparing them to the extent of IS/OS junction loss on the corresponding individual B-scans. Another limitation of the current study was that comparison with NIR-AF imaging was possible only in a small subgroup of patients. Despite these limitations and the small sample size used in this study, we demonstrated that the wide-field en face slab approach has the potential to be a useful clinical tool for detecting and monitoring changes in RPE atrophy, IS/OS junction loss, and fleck distribution in patients with STGD1. In addition, a comparison between en face and SW-AF results has contributed to our understanding of the spatial and intraretinal location of flecks, a prominent clinical feature of STGD1.

Acknowledgments

The authors thank Juan Reynaud for his help with the special purpose software used to generate the en face slab images. Supported in part by grants from the National Eye Institute/NIH (EY09076, EY019861), grants from Topcon, Inc., and a grant from Research to Prevent Blindness to the Department of Ophthalmology, Columbia University.

Disclosure: V.C. Greenstein, None; J. Nunez, None; W. Lee, None; K. Schuch, None; B. Fortune, None; S.H. Tsang, None; R. Allikmets, None; J.R. Sparrow, None; D.C. Hood, Topcon, Inc. (F, C), Heidelberg Engineering (F, C)

References

22. Sisk RA, Leng T. Multimodal imaging and multifocal electroretinography demonstrate autosomal recessive Stargardt...


