The Adenosine Receptor Antagonist, 7-Methylxanthine, Alters Emmetropizing Responses in Infant Macaques

Li-Fang Hung,1,2 Baskar Arumugam,1,2 Lisa Ostrin,1 Nimesh Patel,1 Klaus Trier,3 Monica Jong,2,4 and Earl L. Smith III1,2

1College of Optometry, University of Houston, Houston, Texas, United States
2Brien Holden Vision Institute, Sydney, Australia
3Trier Research Laboratories, Hellerup, Denmark
4Department of Optometry and Vision Science, University of New South Wales, Kensington, New South Wales, Australia

Correspondence: Earl L. Smith III, University of Houston, College of Optometry, 4901 Calhoun Road, 505 J Armistead Building, Houston, TX 77204-2020, USA; esmith@uh.edu.

Submitted: June 1, 2017
Accepted: December 3, 2017
Citation: Hung LF, Arumugam B, Ostrin L, Patel N, Trier K, Jong M, Smith EL III. The adenosine receptor antagonist, 7-methylxanthine, alters emmetropizing responses in infant macaques. Invest Ophthalmol Vis Sci. 2018;59:472–486. https://doi.org/10.1167/iovs.17-22337

Purpose. Previous studies suggest that the adenosine receptor antagonist, 7-methylxanthine (7-MX), retards myopia progression. Our aim was to determine whether 7-MX alters the compensating refractive changes produced by defocus in rhesus monkeys.

Methods. Starting at age 3 weeks, monkeys were reared with −3 diopter (D; n = 10; 7-MX −3D/pl) or +3D (n = 6; 7-MX +3D/pl) spectacles over their treated eyes and zero-powered lenses over their fellow eyes. In addition, they were given 100 mg/kg of 7-MX orally twice daily throughout the lens-rearing period (age 147 ± 4 days). Comparison data were obtained from lens-reared controls (−3D/pl, n = 17; +3D/pl, n = 9) and normal monkeys (n = 37) maintained on a standard diet. Refractive status, corneal power, and axial dimensions were assessed biweekly.

Results. The −3D/pl and +3D/pl lens-reared controls developed compensating myopic (−2.10 ± 1.07 D) and hyperopic anisometropias (+1.86 ± 0.54 D), respectively. While the 7-MX +3D/pl monkeys developed hyperopic anisometropias (+1.79 ± 1.11 D) that were similar to those observed in +3D/pl controls, the 7-MX −3D/pl animals did not consistently exhibit compensating myopia in their treated eyes and were on average isometric (+0.35 ± 1.96 D). The median refractive errors for both eyes of the 7-MX −3D/pl (+5.47 D and +4.38 D) and 7-MX +3D/pl (+5.28 and +3.84 D) monkeys were significantly more hyperopic than that for normal monkeys (+2.47 D). These 7-MX-induced hyperopic ametropias were associated with shorter vitreous chambers and thicker choroids.

Conclusions. In primates, 7-MX reduced the axial myopia produced by hyperopic defocus, augmented hyperopic shifts in response to myopic defocus, and induced hyperopia in control eyes. The results suggest that 7-MX has therapeutic potential in efforts to slow myopia progression.

Keywords: myopia, hyperopia, emmetropization, 7-methylxanthine, adenosine receptors
has not been firmly established in either children or animals (Benavente-Perez A, et al. IOVS 2017;58:ARVO E-Abstract 5466; McFadden SA. IOVS 2017;58:ARVO E-Abstract 5467).

The nonselective adenosine receptor antagonist, 7-methylxanthine (7-MX), may provide an additional pharmaceutical treatment strategy for myopia. The initial interest in 7-MX as a potential therapeutic agent to slow myopia progression emerged from the observation that in contrast to the reduction in scleral collagen content and the decrease in collagen fibril diameter associated with myopic axial elongation, oral 7-MX increased the collagen-related amino acid content, the collagen fibril diameter, and the thickness of the posterior sclera in rabbits, presumably strengthening the sclera and reducing the potential for axial elongation. A subsequent pilot trial found that 7-MX was moderately effective in slowing axial elongation in myopic children. However, the sites and mechanisms of action of 7-MX are unknown.

Establishing an appropriate animal model for investigating the antmyopia effects of adenosine receptor antagonists is a critical step in refining key variables for clinical trials, evaluating safety issues, and identifying how adenosine antagonists influence refractive development. To date the observed effects of adenosine receptor antagonists on refractive development in animals have not been consistent. For instance in rabbits and guinea pigs oral 7-MX reduced the degree of form-deprivation myopia by 18% and 46%, respectively (measured relative to fellow control eyes). However, in chickens, 7-MX did not significantly reduce the degree of form-deprivation myopia and only reduced the degree of myopia produced by hyperopic defocus by 23% (Wang K, et al. IOVS 2014;55:ARVO E-Abstract 3040). The purpose of this investigation was to characterize the effects of 7-MX on normal refractive development and vision-induced ametropias in infant rhesus monkeys.

We chose to investigate 7-MX because it does not have the arousal effects of other adenosine antagonists, such as caffeine, it has low toxicity, no carcinogenic effects, and there have been no reported adverse side effects in children in the previous pilot trials (Trier K, et al. IOVS 2017;58:ARVO E-Abstract 2385). In addition, 7-MX is a common and well-tolerated dietary ingredient. For instance, cacao is one of the most common dietary sources of 7-MX. Although cacao contains only minor amounts of 7-MX, it contains relatively large amounts of theobromine of which 7-MX is the main metabolite. For example, 50 g of dark chocolate contain 250 to 375 mg of theobromine. In humans, 36% of a dose of theobromine is excreted in the urine as 7-MX and, at any given point in time, the serum concentration of 7-MX is approximately 10% of that of theobromine.

Rhesus monkeys were used in this study because the resulting data should be directly applicable to humans. Previous studies have shown a close correspondence, both qualitative and quantitative, between humans and macaques in the course of emmetropization, the changes in ocular components that occur during normal development, and the alterations in ocular components that are associated with ametropias, and the effects of visual experience on refractive development.

**Materials and Methods**

**Subjects**

The primary subjects were 16 rhesus monkeys (Macaca mulatta) that were obtained at 2 to 3 weeks of age and reared in our nonhuman primate nursery. The ambient lighting was provided by fluorescent lights (correlated color temperature = 5500 K Philips TL75; Philips Lighting, Somersert, NJ, USA) maintained on a 12-hour light/12-hour dark cycle with the lights-on cycle starting at 7 AM. The average cage-level illuminance during the daily light cycle was 480 lux (range, 342-688 lux; for husbandry details see Smith and Hung). Beginning at 24 ± 2 days of age, the experimental animals were fitted with light-weight goggles that secured either −3 D (n = 10) or +3 D spectacle lenses (n = 6) in front of their treated eyes and clear, zero-powered (“plano”) lenses in front of their fellow eyes, providing horizontal monocular and binocular fields of view of 80° and 62°, respectively, and an 87° vertical field. Except for brief periods needed for routine cleaning and maintenance, the lenses were worn continuously until 147 ± 4 days of age. Imposed defocus of ±3 D is well within the operating range of the emmetropization process in normal infant monkeys and the lens-rearing period was sufficiently long to ensure complete compensating growth for the degrees of imposed defocus.

In addition to receiving the standard dietary regimen for infant monkeys, throughout the lens-rearing period, the experimental infants were fed by mouth 100 mg/kg of pharmaceutical grade 7-MX (V.B. Medicare Pvt. Ltd, Karnataka, India). The 7-MX was suspended in a small volume of infant formula or apple juice and administered orally twice daily (at 7:30 AM and 6:30 PM). Care was taken to ensure that the animals ingested all of the 7-MX prior to their normal feedings. Comparisons across previous animal studies suggested that lower dosages of 7-MX might not be effective in reducing vision-induced myopia (see the Discussion). Consequently, to reduce the potential for false negative results, we deliberately used a dosage of 7-MX that was approximately 3 times higher than that employed in any previous animal study and that was at least 6.5 times higher than that employed in earlier human studies. 7-MX has a relatively short half-life in the blood stream (~200 minutes in adult humans and 1 hour in rabbits). Consequently, the drug was administered twice each day to increase the daily duration of the drug exposure.

Comparison data, some of which were reported in previous publications, were obtained from infant monkeys that were reared with monocular −3 D (n = 17) or +3 D (n = 9) treatment lenses. The goggles, the control lenses for the fellow eyes, and the onset and duration of lens wear for the lens-reared controls were identical to those for the 7-MX-treated monkeys. Control data were also available for 37 infant monkeys that were reared with unrestricted vision. The rearing procedures, general husbandry protocols, and biometric measurement methods for the lens-reared and normal control animals were identical to those for the 7-MX-treated monkeys, except that they were not fed the adenosine receptor antagonist.

**Ocular Biometry**

The procedural details for measuring the eye’s refractive status, corneal power, and axial dimensions have been described previously. Briefly, the monkeys were anesthetized (intramuscular injection: ketamine hydrochloride, 15–20 mg/kg, and acepromazine maleate, 0.15–0.2 mg/kg; topical: 0.5% tetracaine hydrochloride) and cyclopiaegia was induced by the instillation of 1% tropicamide 25 and 20 minutes prior to obtaining the measurements. The refractive state of each eye was measured independently by two experienced investigators using a streak retinoscope and averaged using matrix notation. An eye’s refractive error was defined as the spherical-equivalent, spectacle-plane refractive correction (95% limits of agreement = ±0.60 D). The anterior radius of curvature of the cornea was measured using a hand-held probe.
keratometer (Alcon Auto-keratometer; Alcon, Inc., St. Louis, MO, USA) or a corneal video topographer (EyeSys 2000; EyeSys vision, Inc., Houston, TX, USA) when the corneal power exceeded the measurement range of the keratometer. Three readings were taken with the hand-held keratometer and averaged to calculate the central corneal power using an assumed refractive index of 1.3375 (95% limits of agreement = ±0.49 to −0.37 D for mean corneal power). Ocular dimensions were measured by A-scan ultrasonography using a 13-MHZ transducer (Image 2000; Mentor, Norwell, MA, USA); 10 separate measurements were averaged (95% limits of agreement for vitreous chamber depth = ±0.05 mm). The initial biometric measures were obtained at ages corresponding to the start of lens wear and every 2 weeks throughout the observation period.

For the 7-MX–treated monkeys and 6 untreated normal monkeys, choroidal thickness was measured with spectral-domain optical coherence tomography (SD-OCT; Spectralis, Heidelberg, Germany), which has an axial resolution of 3 μm per pixel, approximately an order of magnitude finer than that obtained using A-scan ultrasonography. For imaging, the animal’s head was stabilized using a 5-way positioner (X, Y, Z, tip, and tilt) and gas permeable contact lenses were inserted to ensure good optical clarity. All scans were conducted between 9 and 11 AM to avoid potential confounding effects of diurnal variations in choroidal thickness. The OCT’s scan pattern (30° × 25°) was centered and focused on the fovea and the instrument’s enhanced depth imaging mode was used to improve the visibility of the choroidal-scleral border. Thirty-one horizontal scans were obtained using the instrument’s highest resolution protocol resulting in B-scan images of 1536 × 496 pixels; only scans with a quality index of 20 dB or higher were analyzed. The instrument’s auto rescan feature was employed to track anatomic features to ensure that all subsequent scans were performed at the same retinal location as the baseline measurements. The scan data were exported and analyzed using custom Matlab software (MathWorks, Natick, MA, USA), which further enhanced the visibility of the choroidal-scleral interface. An experienced, masked observer manually segmented each scan to identify Bruch’s membrane and the choroidal-scleral interface. The center of the fovea was identified as the deepest point in the foveal pit observed in the central scans. Our primary measure was subfoveal choroidal thickness, defined as the average thickness across 1.8- to 2.1-mm long segments centered on the foveal pit, after correction for lateral magnification. The 95% limits of agreement for between session comparisons in controls was −10.4 to +5.9 μm. The initial choroidal thickness measures were obtained at ages corresponding to the onset of lens wear; subsequent measures were obtained at several time points during the treatment period.

All rearing and experimental procedures were reviewed and approved by the University of Houston’s Institutional Animal Care and Use Committee and were in compliance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Statistical Methods

Statistical analyses were performed using Minitab (Release 16.2.4; Minitab, Inc., State College, PA, USA) and Super ANOVA software (Abacus Concepts, Inc., Berkeley, CA, USA). At ages corresponding to the end of the treatment period, the distribution of refractive errors in normal monkeys is leptokurtic. Consequently, nonparametric Mann-Whitney U tests were used to compare the median end-of-treatment refractive errors between pairs of subject groups. Mixed-design ANOVAs, followed by unpaired t-tests with Greenhouse-Geisser (G-G) corrections for multiple testing, were used to compare the changes in refractive error, anisometropia, and choroidal thickness between groups that occurred during the lens-rearing period. Paired Student’s t-tests were employed to examine the interocular differences in ocular parameters within a given subject group. One-way ANOVAs were used to examine between-group differences in ocular parameters at ages corresponding to the start and end of lens wear. The relationship between refractive error and choroidal thickness and the relationship between refractive error and the ratio between vitreous chamber depth and corneal radius were characterized using linear regression analyses.

Results

There was no evidence that the 7-MX treatment regimen produced any adverse side effects. During our routine biometric measures, we did not observe any abnormal ocular signs associated with the 7-MX treatments. The behaviors of the 7-MX–treated monkeys were observed multiple times each day during the treatment period and there were no discernible differences between the 7-MX monkeys and the monkeys maintained on a standard diet. Over the course of the treatment period, weight gains were similar in the control and 7-MX–treated monkeys and complete blood counts and basic metabolic analyses were performed. Although infant monkeys in all groups showed systematic differences in comparisons to adults (e.g., slightly lower total protein levels and lower numbers and percentages of neutrophils), there were no systematic differences in the blood analyses between the control and 7-MX–treated monkeys. There were no interocular differences in IOP in either the treated or control animals (P = 0.56–0.93). The average and range of IOPs in the fellow eyes of the 7-MX monkeys (11.4 ± 1.6 and 8–14 mm Hg) measured near the end of the lens-rearing period were comparable to those in age-matched control animals (12.3 ± 2.6 mm Hg, P = 0.22; 9–17 mm Hg). The retinas of the 7-MX monkeys appeared normal during ophthalmoscopy and although the thickness of the various retinal layers were not quantified, visual inspection of the cross-sectional OCT images obtained at the end of the treatment period did not reveal any obvious alterations in retinal anatomy in either the lens-treated or fellow eyes of the 7-MX monkeys.

Baseline refractive and biometric measures for the 7-MX–treated animals are provided in Table 1. At the start of the lens-rearing period, the infant monkeys in the 7-MX groups exhibited the moderate degrees of hyperopia that are characteristic of age-matched normal monkeys and the corneal powers and axial dimensions in the 7-MX monkeys were similar to those in the lens-reared controls and normal monkeys (P = 0.46–2.01; P = 0.10–0.76; see Table 1). The refractive errors and ocular dimensions of the two eyes of the 7-MX monkeys were also well matched. There were no significant interocular differences in either 7-MX group in the average refractive errors, corneal powers, anterior chamber depths, lens thicknesses, or vitreous chamber depths (P = 0.09–0.95).

The effects of −3 D of optically imposed anisometropia on refractive development for representative lens-reared controls are illustrated in the top row of Figure 1. Spherical equivalent ametropias are plotted as a function of age for the treated (red symbols) and control eyes (white symbols) for five of 17
### Table 1. Average (±SD) Baseline Refractive and Biometric Measures

<table>
<thead>
<tr>
<th>Group</th>
<th>Refractive Error, D</th>
<th>Corneal Power, D</th>
<th>Anterior Chamber Depth, mm</th>
<th>Lens Thickness, mm</th>
<th>Vitreous Chamber Depth, mm</th>
</tr>
</thead>
<tbody>
<tr>
<td>Treated eye</td>
<td>4.36 ± 0.88</td>
<td>61.42 ± 1.35</td>
<td>2.49 ± 0.14</td>
<td>3.68 ± 0.06</td>
<td>8.43 ± 0.31</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>4.42 ± 0.97</td>
<td>61.28 ± 1.23</td>
<td>2.50 ± 0.15</td>
<td>3.68 ± 0.10</td>
<td>8.41 ± 0.31</td>
</tr>
<tr>
<td>N = 10</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>3.78 ± 1.15</td>
<td>60.28 ± 1.52</td>
<td>2.47 ± 0.07</td>
<td>3.81 ± 0.14</td>
<td>8.72 ± 0.19</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>3.66 ± 1.22</td>
<td>60.11 ± 1.50</td>
<td>2.48 ± 0.10</td>
<td>3.80 ± 0.13</td>
<td>8.76 ± 0.23</td>
</tr>
<tr>
<td>N = 6</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>3.91 ± 1.42</td>
<td>61.48 ± 1.31</td>
<td>2.50 ± 0.13</td>
<td>3.67 ± 0.17</td>
<td>8.61 ± 0.35</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>3.80 ± 1.15</td>
<td>61.32 ± 1.24</td>
<td>2.51 ± 0.15</td>
<td>3.66 ± 0.15</td>
<td>8.62 ± 0.35</td>
</tr>
<tr>
<td>N = 17</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>4.47 ± 1.31</td>
<td>61.48 ± 1.31</td>
<td>2.43 ± 0.12</td>
<td>3.62 ± 0.22</td>
<td>8.41 ± 0.33</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>4.57 ± 1.27</td>
<td>61.32 ± 1.24</td>
<td>2.59 ± 0.18</td>
<td>3.60 ± 0.18</td>
<td>8.38 ± 0.33</td>
</tr>
<tr>
<td>N = 9</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Right eye</td>
<td>3.95 ± 1.82</td>
<td>61.67 ± 1.91</td>
<td>2.58 ± 0.19</td>
<td>3.69 ± 0.22</td>
<td>8.63 ± 0.30</td>
</tr>
<tr>
<td>Left eye</td>
<td>3.93 ± 1.74</td>
<td>61.74 ± 1.81</td>
<td>2.57 ± 0.21</td>
<td>3.70 ± 0.21</td>
<td>8.63 ± 0.30</td>
</tr>
<tr>
<td>N = 37</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

There were no statistically significant interocular or between group difference at the start of the observation period.

### Figure 1. Spherical-Equivalent, Spectacle-Plane Refractive Corrections Plotted as a Function of Age for the Treated (filled symbols) and Fellow Eyes (open symbols) of Representative Lens-Rared Controls (top row) and 7-MX–treated Monkeys (bottom row). All of the animals were reared with −3.0 D lenses in front of their treated eyes and plano lenses in front of their fellow eyes. The thin gray lines in each plot represent data for the right eyes of the 37 normal control monkeys. The panels in each row are arranged from left to right according to the maximum degree of myopic anisometropia at the end of the lens-rearing period. The monkeys represented in panels (A, F and E, J) exhibited the smallest and largest degrees of myopic anisometropia, respectively. The monkeys in panels (C, H) exhibited the median degree of myopic anisometropia in their respective group.
control animals reared with $-3 \, \text{D}$ lenses in front of their treated eyes and plano lenses in front of their fellow eyes. These animals, which were selected to illustrate the range of compensating responses, are ordered in Figures 1A to 1E on the basis of the degree of myopic anisometropia exhibited at the end of the treatment period from the smallest ($\pm 0.38 \, \text{D}$) to largest degrees of compensating anisometropia ($\pm 3.5 \, \text{D}$). The animal in Figure 1C exhibited the median anisometropia for the group ($\pm 2.31 \, \text{D}$). The key point of the data included in the top row of Figure 1 is that $\pm 3 \, \text{D}$ of imposed monocular hyperopic defocus consistently produced relative myopic shifts in the treated eyes resulting in a compensating myopic anisometropia. Only one of 17 $\pm 3 \, \text{D}/\text{pl}$ control animals failed to manifest a myopic anisometropia of at least 1.0 D after approximately 100 days of age.

As illustrated in the bottom row of Figure 1, which shows longitudinal treated- (blue symbols) and control eye ametropias (white symbols) for five of 10 $\pm 3 \, \text{D}$ control monkeys, oral administration of 7-MX greatly reduced the likelihood that an animal would compensate for the imposed monocular hyperopic defocus consistently produced relative myopic shifts in the treated eyes resulting in a compensating myopic anisometropia. Only one of 17 $\pm 3 \, \text{D}/\text{pl}$ control animals failed to manifest a myopic anisometropia of at least 1.0 D after approximately 100 days of age.

As illustrated in the bottom row of Figure 1, which shows longitudinal treated- (blue symbols) and control eye ametropias (white symbols) for five of 10 $\pm 3 \, \text{D}$/pl control monkeys, oral administration of 7-MX greatly reduced the likelihood that an animal would compensate for the imposed monocular hyperopic defocus consistently produced relative myopic shifts in the treated eyes resulting in a compensating myopic anisometropia. Only one of 17 $\pm 3 \, \text{D}/\text{pl}$ control animals failed to manifest a myopic anisometropia of at least 1.0 D after approximately 100 days of age.

As illustrated in Figure 2, which shows data for five representative $\pm 3 \, \text{D}/\text{pl}$ controls, rearing animals with $\pm 3 \, \text{D}$ of optically imposed anisometropia consistently produced compensating hyperopic anisometropias (top row). All nine of the $\pm 3 \, \text{D}$ lens-reared controls developed relative hyperopic refractions in their treated eyes (red symbols). At the end of the lens-rearing period, the degree of anisometropia varied from $\pm 1.25 \, \text{D}$ (Fig. 2A) to $\pm 2.25 \, \text{D}$ (Fig. 3E). Administration of 7-MX did not prevent compensating hyperopic shifts in the treated eye’s refraction (Fig. 2, bottom row). During the lens-rearing period, all of the 7-MX $\pm 3 \, \text{D}/\text{pl}$ monkeys exhibited absolute hyperopic shifts in their treated eyes (blue symbols). At the end of the lens-rearing period, the treated eyes of these 7-MX monkeys were substantially more hyperopic than the average normal monkey (thick solid lines) and five of six 7-MX $\pm 3 \, \text{D}/\text{pl}$ animals showed obvious compensating hyperopic anisometropias. The only animal that failed to exhibit a compensating hyperopic anisometropia developed substantial hyperopic errors in both eyes (Fig. 2F). The effects of 7-MX on compensation for the optically imposed anisometropia are summarized in Figure 3. In comparison to normal monkeys, the lens-reared controls exhibited consistent compensating anisometric changes in
response to either \( -3 \) or \( +3 \) D of imposed monocular defocus (\( -3 \)D/pl, \( F = 33.1, P = 0.0001; +3 \)D/pl, \( F = 29.7, P = 0.0001 \)). At the end of the lens-rearing period, 16 of 17 \( -3 \)D/pl control animals manifested myopic anisometropias that were more than 2 SDs below the mean for normal monkeys (Fig. 3A); the average ametropias for the treated eyes were significantly less hyperopic/more myopic than their fellow control eyes (\( +0.68 \pm 1.77 \) vs. \( +2.78 \pm 1.20 \) D; \( P < 0.001 \)). Similarly, all nine of the \( +3 \)D/pl control animals developed compensating hyperopic anisometropias that were larger than any anisometropias observed in normal monkeys (Fig. 3C); the average treated eye ametropias were significantly more hyperopic than their fellow control eyes (\( +4.77 \pm 0.91 \) vs. \( +2.91 \pm 1.03 \) D; \( P < 0.001 \)).

Administration of 7-MX eliminated the consistent myopic anisometropia normally produced by \( -3 \) D of imposed monocular defocus. At the end of the lens-rearing period, only two of 10 7-MX \( -3 \)D/pl monkeys had myopic anisometropias (Fig. 3B). The average treated and fellow eye ametropias of the 7-MX \( -3 \)D/pl animals were similar (\( +4.89 \pm 3.04 \) vs. \( +4.54 \pm 2.11 \) D; \( P = 0.59 \)). The temporal pattern of anisometropia for the 7-MX \( -3 \)D/pl monkeys was significantly different from that for the \( -3 \)D/pl controls (\( F = 7.7, P = 0.0005 \)), and at the end of the treatment period the average anisometropia was significantly smaller and more hyperopic than the compensating myopic anisometropia observed in the \( -3 \)D/pl controls (\( +0.34 \pm 1.96 \) vs. \( -2.10 \pm 1.01 \) D; \( P = 0.003 \)). In contrast, compensation for \( +3 \) D of imposed monocular defocus was robust in five of six 7-MX \( +3 \)D/pl monkeys (Fig. 3D). Although one 7-MX \( +3 \)D/pl monkey was essentially isometropic at the end of the lens-rearing period, on average the treated eyes of these animals were significantly more hyperopic than their fellow eyes (\( +6.10 \pm 1.71 \) vs. \( +4.31 \pm 1.97 \) D; \( P = 0.01 \)). Over the course of the treatment period, the pattern of anisometropia for the 7-MX \( +3 \)D/pl animals was not different from that for the \( +3 \)D/pl controls (\( F = 0.72, P = 0.55 \)) and the final degrees of anisometropia were comparable (\( 1.79 \pm 1.11 \) vs. \( +1.86 \pm 0.54 \) D; \( P = 0.81 \)).

In comparison to normal monkeys, 7-MX changed the course of emmetropization in both the treated and fellow eyes of the lens-reared monkeys (\( F = 3.3–17.4, P = 0.05–0.0001 \)). This point is emphasized in Figures 4A and 4B, which show the longitudinal changes in refractive error for the treated (blue symbols) and fellow eyes (white symbols) of the 7-MX \( -3 \)D/pl and 7-MX \( +3 \)D/pl monkeys, respectively. At ages corresponding to the start of the lens-rearing period, the normal monkeys were typically moderately hyperopic and, as illustrated by their median data (gray squares), exhibited systematic reductions in hyperopia during the course of emmetropization. In comparison, only one 7-MX \( -3 \)D/pl monkey, the animal that exhibited clear evidence of myopic anisometropic compensation (Fig. 1J), showed larger reductions in hyperopia than the median normal animal. In the 7-MX \( -3 \)D/pl group, absolute increases in
The data normalized to the refractive errors at the start of the treatment period (the first symbol in each function). The gray squares represent the median changes in refractive error for normal age-matched control animals. The stippled area demarcates the 25% and 75% limits for the normal animals. (C) Ametropias obtained at ages corresponding to the end of the lens-rearing period for individual animals. The right and left eyes of the normal monkeys are represented by open symbols. For the lens-reared animals, the filled and open symbols represent the treated and control eyes, respectively. The short solid lines connect the data for the two eyes of a given animal. The dashed horizontal line represents the right eye median for normal monkeys.

Hyperopia were observed in seven treated and five fellow eyes. In the 7-MX +3D/pl animals, all of the treated eyes and four of six fellow eyes showed absolute increases in hyperopia. As a result, the average changes in refractive error that took place over the course of the lens-rearing period for both the treated and fellow eyes of the 7-MX−3D/pl (+0.53 ± 2.65 D and +0.13 ± 1.47 D) and the 7-MX+3D/pl monkeys (+2.32 ± 1.06 D and +0.66 ± 1.13 D) were significantly more hyperopic than those found in normal, age-matched monkeys (−1.47 ± 1.70 D; F = 5.1–27.9, P = 0.0001–0.03). Moreover, the average refractive-error changes during the treatment period in the 7-MX monkeys were more hyperopic than those observed in the treated and fellow eyes of the lens-reared controls (−3D controls: treated eyes, −3.23 ± 1.09 D, P = 0.001 and fellow eyes, −1.02 ± 0.65, P = 0.04; +3D controls: treated eyes, +0.30 ± 1.36 D, P = 0.004 and fellow eyes, −1.66 ± 1.45, P = 0.002).

A comparison of the end of treatment ametropias between the 7-MX monkeys and the control and normal monkeys emphasizes how 7-MX altered the course of emmetropization in both the treated and fellow eyes (Fig. 4C). At the end of lens-rearing period, the median refractive errors for both the treated and fellow control eyes of the 7-MX−3D/pl (+5.47 and +4.28 D) and the 7-MX+3D/pl monkeys (+5.28 and +3.84 D) were significantly more hyperopic than for normal age-matched monkeys (+2.47 D; P = 0.0003–0.01). The median end of treatment ametropias for the treated and fellow eyes of the 7-MX−3D/pl monkeys were also more hyperopic than the treated− (+0.48 D, P = 0.002) and fellow-eye medians (+2.75 D, P = 0.05) for the −3D/pl lens-reared controls. Similarly, at the end of the treatment period, there was a trend for the median ametropias of the treated and fellow eyes of the 7-MX+3D/pl to be more hyperopic than those of the +3D/pl controls (treated eyes: +4.50 D, P = 0.08; fellow eyes: +2.88 D, P = 0.22).

Figure 5 shows the end of treatment corneal powers (Fig. 5A), anterior chamber depths (Fig. 5B), and lens thicknesses (Fig. 5C) for both eyes of individual animals in all five subject groups (see Table 2). Inspection of Figure 5 reveals that neither the lens-rearing procedures nor the administration of 7-MX altered these ocular dimensions. For the lens-reared controls and the 7-MX animals, there were no significant interocular differences in corneal power (T = −1.87 to 0.71, P = 0.10–0.49), anterior chamber depth (T = −0.50 to 1.49, P = 0.18–0.84), or lens thickness (T = −1.66 to 0.42, P = 0.14–0.69).

With the exception of the corneal powers of one monkey in the −3 D control group, all of the biometric measures for the lens-reared controls and the 7-MX–treated monkeys fell within 2 SDs of the average values for normal monkeys. Moreover, 1-way ANOVAs demonstrated that at the end of the treatment period there were no between-group differences in absolute corneal power (F = 2.01, P = 0.10), anterior chamber depth (F = 0.20, P = 0.94), or lens thickness (F = 1.19, P = 0.32).

The axial basis of the refractive-error alterations produced by the lens-rearing procedures and the 7-MX treatment is illustrated in Figure 6. Figure 6A shows the changes in vitreous chamber depth that took place in both eyes of individual monkeys. There are two key points to Figure 6A. First, the three subject groups that exhibited significant degrees of anisometropia at the end of the treatment period also exhibited interocular differences in vitreous chamber elongation. In particular, the treated eyes of the −3D/pl controls increased more in vitreous chamber depth than their fellow control eyes (1.82 ± 0.31 vs. 1.45 ± 0.25 mm, F = 3.96, P = 0.001). On the other hand, compared with that in their fellow eyes, vitreous chamber elongation was significantly retarded in the treated eyes of the +3D/pl controls (1.10 ± 0.39 vs. 1.45 ± 0.37 mm, T = −7.79, P < 0.001) and the 7-MX+3D/pl monkeys (0.85 ± 0.24 vs. 1.25 ± 0.24 mm, T = −5.25, P = 0.003). The 7-MX−3D/pl monkeys did not develop significant degrees of anisometropia during the lens-rearing period and there were no systematic interocular differences in vitreous chamber elongation between their treated and fellow eyes (1.08 ± 0.54 vs. 1.05 ± 0.32 mm, T = 0.31, P = 0.77).

The second key point illustrated in Figure 6A is that the relative hyperopic refractive errors found in both eyes of the 7-MX−3D/pl monkeys were associated with significantly smaller than normal amounts of vitreous chamber elongation. For example, the fellow eyes of the 7-MX−3D/pl animals showed...
lower degrees of vitreous chamber elongation in comparison to normal monkeys (1.20 ± 0.32 mm; \( T = -1.78, P = 0.05 \)) and both the treated (\( T = -1.26, P = 0.001 \)) and fellow eyes (\( T = -3.15, P = 0.006 \)) of the 7-MX −3D/pl monkeys exhibited smaller degrees of vitreous chamber elongation than that observed in the −3D/pl controls. Similarly, the increases in vitreous chamber depth in the treated (\( T = -1.43, P = 0.18 \)) and fellow eyes (\( T = -1.29, P = 0.23 \)) of the 7-MX +3D/pl monkeys were smaller than those in the +3D/pl controls, but these differences were not significant.

The end of treatment ametropias are plotted as a function of the ratio between the final absolute refractive errors and the VC/CR depth. In this respect, the highly significant correlation between the final absolute refractive errors and the VC/CR ratio (\( r^2 = 0.87, P < 0.0001 \)) demonstrates that the treatment-related, between-subject differences in refractive error were due to the effects of the lens-rearing procedures and the 7-MX treatments on vitreous chamber elongation.

The 7-MX treatment regimen produced substantial increases in choroidal thickness. In Figures 7A and 7B the longitudinal changes in choroidal thickness are plotted for the treated (blue symbols) and fellow eyes (white symbols) of the 7-MX −3D/pl and 7-MX +3D/pl subjects, respectively. For reference the solid gray symbols in each plot show the average choroidal thickness changes in the left eyes of six normal untreated monkeys. Over ages that corresponded to the lens-rearing period, the normal monkeys exhibited, on average, a small increase in choroidal thickness (7.6 ± 3.4 μm). Both the treated and fellow eyes of

### Table 2. Average (±SD) End of Treatment Refractive and Biometric Measures

<table>
<thead>
<tr>
<th>Group</th>
<th>Refractive Error, D</th>
<th>Corneal Power, D</th>
<th>Anterior Chamber Depth, mm</th>
<th>Lens Thickness, mm</th>
<th>Vitreous Chamber Depth, mm</th>
</tr>
</thead>
<tbody>
<tr>
<td>7-MX −3D/pl</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>+4.89 ± 3.04*</td>
<td>55.97 ± 1.29</td>
<td>3.05 ± 0.16</td>
<td>3.66 ± 0.10</td>
<td>9.51 ± 0.76*</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>+4.54 ± 2.11*</td>
<td>56.21 ± 1.37</td>
<td>3.05 ± 0.17</td>
<td>3.68 ± 0.11</td>
<td>9.46 ± 0.49*</td>
</tr>
<tr>
<td>N = 10</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>7-MX +3D/pl</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>+6.10 ± 1.71*</td>
<td>53.92 ± 1.17</td>
<td>3.08 ± 0.14</td>
<td>3.73 ± 0.17</td>
<td>9.57 ± 0.40*</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>+4.31 ± 1.97*</td>
<td>54.11 ± 1.05</td>
<td>3.08 ± 0.15</td>
<td>3.72 ± 0.17</td>
<td>10.01 ± 0.44</td>
</tr>
<tr>
<td>N = 6</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>−3D/pl control</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>+0.68 ± 1.77*</td>
<td>55.16 ± 1.49</td>
<td>3.13 ± 0.14</td>
<td>3.64 ± 0.11</td>
<td>10.43 ± 0.55*</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>+2.78 ± 1.20</td>
<td>55.08 ± 1.56</td>
<td>3.11 ± 0.12</td>
<td>3.65 ± 0.13</td>
<td>10.05 ± 0.46</td>
</tr>
<tr>
<td>N = 17</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>+3D/pl control</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Treated eye</td>
<td>+4.77 ± 0.91*</td>
<td>55.80 ± 1.53</td>
<td>3.13 ± 0.15</td>
<td>3.54 ± 0.13</td>
<td>9.51 ± 0.51*</td>
</tr>
<tr>
<td>Fellow eye</td>
<td>+2.91 ± 1.03</td>
<td>56.13 ± 1.14</td>
<td>3.03 ± 0.09</td>
<td>3.56 ± 0.14</td>
<td>9.83 ± 0.31</td>
</tr>
<tr>
<td>N = 9</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Normals</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Right eye</td>
<td>+2.46 ± 1.04</td>
<td>55.88 ± 1.82</td>
<td>3.04 ± 0.19</td>
<td>3.67 ± 0.15</td>
<td>9.83 ± 0.51</td>
</tr>
<tr>
<td>Left eye</td>
<td>+2.48 ± 1.01</td>
<td>55.80 ± 1.77</td>
<td>3.04 ± 0.18</td>
<td>3.68 ± 0.16</td>
<td>9.84 ± 0.32</td>
</tr>
<tr>
<td>N = 37</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Significantly different from normal monkeys (\( P < 0.05 \)).  
† Significant interocular difference (\( P < 0.05 \)).
the 7-MX monkeys exhibited significantly larger increases in choroidal thickness ($F = 2.6–9.1$, $P = 0.05–0.0002$). At the end of the lens-rearing period, the increases in choroidal thickness for the 7-MX subjects were between 3.2 to 5.1 times larger than those observed in the normal monkeys. These increases in choroidal thickness in the treated eyes of the 7-MX/C03D/pl monkeys are particularly noteworthy because imposed hyperopic defocus has been shown to produce choroidal thinning in control monkeys.83

At the end of the treatment period there were no interocular differences in choroidal thickness between the treated and fellow eyes of the 7-MX/C03D/pl animals ($t = 1.52$, $P = 0.16$). On the other hand, the choroids in the lens-treated eyes of the 7-MXþ3D/pl monkeys were significantly thicker than those in their fellow eyes ($170.1 \pm 26.0$ vs. $163.4 \pm 25.1$; $T = 2.69$, $P = 0.04$). In lens-reared control monkeys, changes in choroidal thickness are significantly correlated with the degree of compensating anisometropia ($2.7 \mu m/D$).87 At the end of the lens-rearing period, the treated eyes of the 7-MXþ3D/pl monkeys were on average approximately 2 D more hyperopic than their fellow eyes and the treated-eye choroids were 6.7–μm thicker. In this respect, the magnitude of the interocular differences in choroidal thickness observed in the 7-MXþ3D/pl animals suggests that the 7-MX did not substantially alter the choroidal thickness changes produced by myopic defocus. However, it is important to note that although the increases in choroidal thickness were in the appropriate direction to contribute to the hyperopic ametropias in both groups of 7-MX–treated monkeys and the hyperopic anisometropias in the 7-MXþ3D/pl monkeys, the largest observed changes in choroidal thickness would account for less than a 0.25 D change in refractive error (i.e., the alterations in axial elongation noted above dominated the observed refractive error changes).

The changes in choroidal thickness, which occurred shortly after the onset of treatment and remained relatively stable throughout the observation period, were positively correlated

![Figure 6](http://tvst.arvojournals.org/)

**Figure 6.** (A) Relative changes in vitreous chamber depth measured over ages corresponding to the lens-rearing period for individual normal monkeys (open diamonds) and the treated (filled symbols) and control eyes (open symbols) of the lens-reared controls (red symbols) and the 7-MX monkeys (blue symbols) reared with −3 D or +3 D of imposed anisometropia. The short solid lines connect the data for the two eyes of a given animal. The solid horizontal line indicates the average for the normal monkeys and the dashed lines represent ±1 SD from the mean. (B) End of treatment ametropias plotted as a function of the vitreous chamber/corneal radius ratios for the eyes of individual lens-reared monkeys. The blue and red symbols represent data from the 7-MX– and control lens-reared monkeys, respectively. The squares and diamonds represent data for the treated eyes that viewed through +3 D and −3 D lenses, respectively. The circles represent data for the fellow control eyes.

![Figure 7](http://tvst.arvojournals.org/)

**Figure 7.** The average (±SEM) relative changes in choroidal thickness for the treated (blue symbols) and fellow control eyes (open symbols) of 7-MX–treated monkeys reared with −3 D (A) and +3 D of imposed anisometropia (B). The solid gray symbols represent data from untreated normal eyes. The asterisks indicate significant differences ($P < 0.05$) between the normal and 7-MX monkeys. (C) Changes in ametropia that took place during the lens-rearing period plotted as a function of the changes in choroidal thickness for individual animals. The gray squares represent untreated normal eyes. The circles and diamonds symbols represent the 7-MXþ3D/pl and 7-MX−3D/pl monkeys, respectively. The treated and fellow eyes of the 7-MX–treated monkeys are represented by the filled and open symbols, respectively. The solid line was determined by linear regression analysis.
with the refractive error changes observed in the 7-MX monkeys (Fig. 7C; $r^2 = 0.28, P = 0.001$). While, as stated above, the absolute changes contributed little to the absolute change in refractive error, the choroidal changes preceded the refractive changes and were predictive of the magnitude of the refractive changes.

**Discussion**

Our main finding was that the oral administration of the nonselective adenosine receptor antagonist, 7-MX, altered vision-dependent emmetropization in infant monkeys in a direction-specific manner. Specifically, the 7-MX regimen produced absolute hyperopic shifts in the refractive errors of both the treated and fellow eyes of the 7-MX monkeys. Moreover, the 7-MX treatments dramatically reduced the compensating axial myopic anisometropias normally produced by imposed monocular hyperopic defocus. However, in contrast, the 7-MX regimen did not prevent, and in fact augmented, the compensating hyperopic refractive errors normally produced by imposed monocular myopic defocus. The alterations in refractive development observed in both eyes of the 7-MX monkeys were axial in nature, being highly correlated with vitreous chamber depth.

**Comparisons Between Different Species**

The effects of 7-MX on vision-induced ocular growth and refractive development in infant monkeys are in some respects qualitatively similar to those observed previously in guinea pigs and rabbits. It was hypothesized that the changes produced by 7-MX would improve the biomechanical properties of the sclera reducing the potential for myopic axial elongation. The observation that all known subtypes of adenosine receptors are found in human scleral fibroblasts supported the idea that adenosine antagonists could directly influence scleral remodeling.

In the fibrous layers of the sclera, particularly in the primate eyes, myopic axial elongation is associated with scleral remodeling that includes a reduction in collagen content, a decrease in collagen fibril diameter, and a decrease in overall thickness of the posterior sclera. These changes alter the mechanical properties of the sclera making it more extensible and more susceptible to myopic axial elongation. The initial interest in adenosine receptor antagonists as potential therapeutic agents to slow myopia progression emerged from the observation that oral administration of either theobromine or 7-MX, both metabolites of caffeine, increased the collagen-related amino acid content, the collagen fibril diameter, and the thickness of the posterior sclera in rabbits. It was hypothesized that the changes produced by 7-MX would improve the biomechanical properties of the sclera reducing the potential for myopic axial elongation. The observation that all known subtypes of adenosine receptors are found in human scleral fibroblasts supported the idea that adenosine antagonists could directly influence scleral remodeling.

The increases in choroidal thickness observed in the 7-MX-treated monkeys do not necessarily rule out the possibility that the primary site of action of 7-MX is the sclera. However, given that the choroid is a critical component in the vision-dependent cascade that mediates normal refractive development, it is possible that 7-MX affects refractive development by influencing or altering the operational properties of this emmetropization cascade. A large body of evidence supports the hypothesis that vision-dependent emmetropization is regulated by local retinal mechanisms, which generate biochemical signals that reflect the eye’s effective refractive state. The signal cascade passes from the inner retina, through the retinal pigment epithelium (RPE) and choroid, and eventually modulates the biochemical and mechanical nature of the sclera in a manner that promotes changes in overall axial length that normally eliminate refractive errors. The effects of 7-MX on refractive development could have come about by influencing this visually driven signal cascade at a number of different locations. All four subtypes of adenosine receptors have been identified in the neural retina, pigment epithelium, choroid, and sclera. At least some of these receptors appear to be involved in emmetropization and the regulation of axial elongation. For instance, the pattern of adenosine receptor expression in different ocular tissues is altered by form deprivation in guinea pigs and genetic deletion of the A2A adenosine receptor in mice promotes axial myopia. Although it is possible our 7-MX treatments indirectly influenced the emmetropization process via nonadenosine receptors (e.g., via off-target actions on purinergic receptors), the known effects of adenosine mechanisms in nonocular tissue suggest that receptor antagonists, such as 7-MX, could influence the retinal components of the signal cascade in a number of ways (e.g., by modulating dopamine or acetylcholine transmission, or altering ocular circadian
7-MX Retards Ocular Axial Elongation

**Clinical Implications**

Our results demonstrate that adenosine receptor antagonists have potential in treatment regimens for preventing or slowing the progression of myopia in children and provide support for the recent clinical trials investigating the efficacy of 7-MX in retarding myopia progression. As a therapeutic agent for treating myopia, 7-MX has a number of positive attributes. Most importantly, 7-MX, which is a common dietary ingredient, has a good safety record in the limited clinical trials conducted to date, with no significant adverse effects noted over treatment periods that were 3 years in duration for some children (Trier K, et al. IOVS 2017;58:ARVO E-Abstract 2385).

In this respect, we did not observe any adverse side effects in the 7-MX–treated monkeys. The results for this study suggest that 7-MX could be beneficial in slowing myopia progression in two ways. First, 7-MX retarded the ability of hyperopic defocus, the presumed stimulus for myopia progression in children, to produce myopia. Second, 7-MX did not interfere with the reduction in axial elongation produced by imposed myopic defocus, and in fact may have exaggerated these optical effects. It has been suggested that defocus signals produce separate “stop” and “go” signals that modulate axial elongation. If this is correct, 7-MX appears to reduce “go” signals and to possibly amplify “stop” signals. Regardless, this second point has practical significance, because it supports the idea that combining optical treatment strategies that impose relative myopic defocus over all or a large part of the visual field with 7-MX would have a greater effect than either 7-MX or the optical treatment strategies alone. Although our results are encouraging, it is important to recognize that the site and mechanism of action of 7-MX are still unknown.

Are the hyperopic shifts, particularly those observed in the fellow control eyes of the 7-MX–treated monkeys, a contraindication for using adenosine antagonists to slow myopia progression in children? For example, would 7-MX produce unwanted hyperopia in treated myopic children? This possibility is unlikely primarily because of the ocular basis for the hyperopic shifts produced by 7-MX. As described above, the hyperopia came about because 7-MX selectively slowed vitreous chamber elongation without altering the normal reduction in the eye’s refracting power that occurred as the cornea and crystalline lens followed their normal growth patterns. In this respect, current treatment regimens for myopia progression, which all slow vitreous chamber elongation without altering the refracting power of the eye, have the potential to produce hyperopic shifts in infant eyes, specifically in eyes in which the cornea and crystalline lens have not reached optical maturity. As shown in Figure 2, optical treatment strategies that imposed myopic defocus can produce hyperopia in infant primates. Similarly, chronic topical atropination can produce axial hyperopia in infant monkeys and infant mice.

It is important to note that the potential for treatment strategies that slow axial elongation to produce hyperopia decreases dramatically with the subject’s age. Once, the cornea and crystalline lens are optically mature, 7-MX, atropine, and the currently employed optical treatment strategies would not be expected to produce hyperopic shifts. In humans, the cornea assumes adult refracting powers as early as 18 to 24 months of age. The greatest changes in lens power occur before 5 years of age with only small, slow changes in lens power occurring after the typical onset ages for juvenile-onset myopia (8–10 years in the United States). As a consequence,
at ages when these antimyopia treatment strategies would be employed, it is highly unlikely that hyperopia would be produced.

With respect to extrapolating our results to clinical treatment strategies, it is also important to note that the dosage of 7-MX that we employed in monkeys was substantially higher than those used in the previous clinical trials involving children. Whereas our animals received 100 mg/kg of 7-MX twice daily, children were prescribed either 400 mg of 7-MX once or twice a day (i.e., 800 mg total). Based on the average weights of 8-26 kg and 13-year-old Danish females (45 kg), the per kilogram dosages of 7-MX used in the previous clinical trials varied from a low of 8.9 to a high of 30.8 mg/kg per day, which were for some children more than 22 times lower than the total per kilogram daily dosages employed in this study. From an efficacy perspective, it will be important to evaluate the effects of the 7-MX dose levels on the antimyopia effects observed in this study. Moreover, although we did not observe any overt behavioral, ocular, or systemic adverse effects related to the 7-MX treatments, the dosages that we employed were higher than those used in any previous animal or human study and we cannot, at this time, rule out subtle toxic effects in either the retina or other ocular structures. Thus, from a safety perspective a thorough examination of the function and structure of the retina, pigment epithelium, choroid, and sclera in 7-MX-treated animals is essential. Nevertheless, the results from this study support further investigations into the efficacy of 7-MX in therapeutic regimens to reduce myopia progression.

Acknowledgments

Supported by National Institutes of Health Grants EY03611 and EY 07551 (Bethesda, MD, USA) and funds from the Brien Holden Vision Institute (Sydney, Australia) and the University of Houston Foundation (Houston, TX, USA).

Disclosure: L.-F. Hung, None; B. Arumugam, None; L. Ostrin, None; N. Patel, None; K. Trier, Theialife (I); P. M. Jong, None; E.L. Smith III, None

References


42. Chia A, Lu QS, Tan D. Five-year clinical trial on atropine for the treatment of myopia 0.5%: myopia control with atropine 0.01% eyelids. *Optom Vis Sci*. 2016;56:391–399.


7-MX Retards Ocular Axial Elongation


100. Oliveira L, Correa-de SA P. Protein kinase A and CatVJ (L-type) channels are common targets to faciltatory adenosine A2A and muscarinic M1 receptors on rat motoneurons. Neurosignals. 2005;14:262–272.


